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Assessing the colloidal properties of engineered nanoparticles in water: case studies from fullerene C₆₀ nanoparticles and carbon nanotubes

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Environmental context. The fate and bioavailability of engineered nanoparticles in natural aquatic systems are strongly influenced by their ability to remain dispersed in water. Consequently, understanding the colloidal properties of engineered nanoparticles through rigorous characterisation of physicochemical properties and measurements of particle stability will allow for a more accurate prediction of their environmental, health, and safety effects in aquatic systems. This review highlights some important techniques suitable for the assessment of the colloidal properties of engineered nanoparticles and discusses some recent findings obtained by using these techniques on two popular carbon-based nanoparticles, fullerene C_{60} and multi-walled carbon nanotubes.

Abstract. The colloidal properties of engineered nanoparticles directly affect their use in a wide variety of applications and also control their environmental fate and mobility. The colloidal stability of engineered nanoparticles depends on their physicochemical properties within the given aqueous medium and is ultimately reflected in the particles' aggregation and deposition behaviour. This review presents some of the key experimental methods that are currently used to probe colloidal properties and quantify engineered nanoparticle stability in water. Case studies from fullerene C_{60} nanoparticles and multi-walled carbon nanotubes illustrate how the characterisation and measurement methods are used to understand and predict nanoparticle fate in aquatic systems. Consideration of the comparisons between these two classes of carbon-based nanoparticles provides useful insights into some major current knowledge gaps while also revealing clues about needed future developments. Key issues to be resolved relate to the nature of near-range surface forces and the origins of surface charge, particularly for the reportedly unmodified or 'pure' carbon-based nanoparticles.

Additional keywords: aggregation, deposition, DLVO, dynamic light scattering, X-ray photoelectron spectroscopy.

Introduction

Over the last decade, nanotechnology has matured into a prominent, interdisciplinary field that influences almost every major branch of science.[1] A fundamental building block of nanotechnology is the engineered nanoparticle (ENP). These particles are by definition of a size between 1 and 100 nm in a least one dimension, [2] and come in a variety of geometries such as spheres, [3] cylinders, [4] and planes. [5] As a subset of traditional colloids (particles between 1 nm and a few micrometres in diameter), nanomaterials are unique because they are on a length scale in which surface and quantum effects become increasingly important. [2] Indeed, these effects often provide ENPs with unusual reactivity as well as novel optical, mechanical, and electrical properties. Compositionally, ENPs may be broadly categorised as being either carbonaceous (e.g. fullerene C₆₀ nanoparticles and carbon nanotubes (CNTs)) or non-carbonaceous (e.g. zero-valent iron nanoparticles, silver nanoparticles, and quantum dots).

The combination of chemical and physical attributes exhibited by ENPs has led to their increased use in commercial applications. [6] This is reflected by the fact that their global market has grown steadily from US\$7.5 billion in 2003 to US\$12.7 billion in 2008. Over the next 5 years, their market value is expected to exceed US\$27 billion. [7,8] Current estimates suggest that more than 800 consumable products, including cell phone batteries, sporting equipment, and cosmetics, already contain ENPs. [9] In addition to these everyday items, work is being conducted to investigate the potential of ENPs in environmental remediation strategies [10–15] and advanced medical procedures. [16,17] For example, the US Environmental Protection Agency (US EPA) recently (2008) released data on 26 sites across North America in which nanomaterials were being used to remove contaminants such as chlorinated solvents from groundwater. [18] In nanomedicine, ENPs are being studied for use in tissue scaffolds, as targeted drug delivery agents, and as components of gene therapy. [16]

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In many applications, the interfacial properties of ENPs (e.g. cohesion and adhesion, wettability and adsorption) are tailored using either covalent or non-covalent surface-modification strategies. For instance, colloidal dispersions of ENPs are commonly desired for industrial applications, but such dispersions can be impossible to prepare with unmodified ENPs owing to the fact that the attractive forces between particles induce aggregation. One approach to overcome this obstacle is to covalently graft functional groups into the ENP surface. Alternatively, surfactants may be physisorbed onto the surface of the ENP in a non-covalent modification process. In either case, interparticle repulsion is increased and dispersion properties are improved. [14,19] Surface modification techniques are also used to deliberately modify other properties (e.g. sorption and wettability) and to create points of attachment for more complex hierarchical nanostructures.[20,21]

As the global market for nanomaterials and nanomaterialcontaining products continues to expand, increasing quantities of ENPs are expected to enter the environment from manufacturing effluents, discarded nanoproducts, or incidental spills. [22,23] The environmental fate, transport, and bioavailability of these ENPs are dependent on their aggregation and deposition behaviour. Comparative toxicity studies have also been conducted to better understand the environmental risks associated with carbonaceous and inorganic nanoparticles in different fluid phase environments. [24–28] In these studies, particular attention has been paid to ENP composition, dispersion state, size distributions, chemical treatments, and surface chemistry, as well as the chemistry of the aquatic environment (e.g. pH, ionic strength, and the concentration of natural organic matter (NOM)). The upshot from these studies is that while some ENPs are toxic to some organisms, the degree of toxicity depends on both the intrinsic properties of the manufactured ENPs and their

physicochemical state in the environment, as also influenced by solution chemistry and the presence of co-contaminants. It is interesting to note that the same variables play a determinant role in the colloidal stability of ENPs.

Thus, it is clear that to develop models that allow us to anticipate and adequately address the environmental risks posed by different ENPs, more detailed information on their behaviour in aquatic environments is required. Structure-property relationships that relate the ENPs' environmental behaviour to their physicochemical properties (e.g. size, shape, and surface chemistry) are also needed. In this review, we will highlight some of the experimental methods currently being used to characterise ENPs and probe their colloidal properties. We then show how several of these techniques have been applied to evaluate the colloidal properties of two important carbonaceous nanoparticles - fullerene C60 nanoparticles and multiwalled carbon nanotubes (MWNTs). To conclude the review, we describe the implications of these findings on general issues of ENP fate and transport and suggest directions for further research.

Colloidal stability

Aggregation

A suspension with superior colloidal stability is one in which the suspended colloidal particles are able to resist aggregation for an indefinite period of time. In this context, aggregation refers to the association of colloidal particles to form larger clusters. [29] The fate and transport of ENPs when released into natural and engineered aquatic systems is largely determined by their aggregation behaviour. Specifically, the rate of ENP aggregation will influence their rate of sedimentation and thus their removal from the aqueous phase.



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Depending on the colloidal stability of the ENPs and the chemistry of the surrounding environment (e.g. pH and electrolyte composition), ENPs can undergo aggregation in solution among themselves before encountering any natural constituents. This process of association among colloidal particles of the same kind is referred to as homoaggregation. In systems containing more than one type of suspended particles, heteroaggregation (or association between particles of different types) can take place either exclusively or simultaneously with homoaggregation. The frequency of homoaggregation and heteroaggregation will depend on the particle concentrations, physical and chemical properties of the particles, and solution chemistry. [30-33] In natural aquatic systems, the fate of ENPs is more likely to be controlled by heteroaggregation between ENPs and naturally occurring colloidal particles because the concentration of naturally occurring colloids will greatly exceed that of ENPs. Nevertheless, homoaggregation is a suitable starting point for investigating the colloidal stability of ENPs because a singlecomponent suspension is a much simpler system to study systematically in the laboratory and to understand conceptually.

Aggregation may be conceptualised as a two-stage process: transport and attachment. [29] First, ENPs and other colloidal particles must be transported towards each other before aggregation can occur. The mechanisms of achieving particle—particle contact can be broadly classified as Brownian diffusion (perikinetic aggregation), fluid motion (orthokinetic aggregation), and differential sedimentation. Further details regarding these transport mechanisms can be found elsewhere. [29] The focus of the present review paper, however, is on the attachment stage of aggregation. The likelihood of a permanent attachment resulting from the approach of two particles is controlled by short-range interparticle forces of interaction, which are in turn dependent on the surface chemistry and composition of the particles as well as the solution chemistry.

Deposition

Another key process that controls the fate and transport of ENPs in natural and engineered aquatic systems is deposition. Deposition refers to the process in which ENPs collide and stick over time to an immobile solid (collector) surface, such as sand, sediments, and rocks. As the solid surface is composed of a different material from the ENPs and can be treated as an infinitely large stationary particle, deposition of ENPs can be considered as an extreme form of heteroaggregation. [29] Deposition can take place in different natural environments, such as surface water and groundwater, and is especially important in systems where solid surfaces for attachment are readily available. Thus, ENP deposition is expected to play a crucial role in influencing the transport of ENPs in groundwater flow through porous media, owing to the vast opportunities for ENP attachment to collector surfaces.

As with aggregation, deposition begins when ENPs are transported to the collector surface. Transport mechanisms for deposition are analogous to those for aggregation and include Brownian diffusion, interception, and sedimentation, with details described elsewhere. [29,34] In the attachment stage, both hydrodynamic effects and colloidal interactions are important factors that will influence the adherence of ENPs to solid surfaces. [29]

Derjaguin-Landau-Verwey-Overbeek (DLVO) theory

The colloidal stability of charged particles dispersed in aqueous solutions is often controlled by their van der Waals and electric double layer interactions. [29,35] The van der Waals interactions between colloidal particles are the result of the interactions between the permanent or induced dipoles within the ENPs and are dependent on the particles' composition, size and geometry. For most particles in aqueous environments, van der Waals interactions are attractive and thus promote aggregation.

Conversely, electric double layer interactions occur owing to charges that reside on the particle surface. The origins of surface charge vary and depend on the surface composition of ENPs. Surface charge is often a consequence of dissociable surface functional groups. [29,35] Some examples of ENPs with surface functional groups include oxidised CNTs, [36] which we will later discuss extensively in this paper, citrate-coated silver nanoparticles,[37,38] and metal oxide nanoparticles such as titanium, [39,40] cerium, [41] and iron oxides. [42] The sign and magnitude of the charge imparted by surface functional groups will depend on their surface site density and solution pH. From a coulombic standpoint, ENPs exhibiting charge of the same sign will repel one another, whereas oppositely charged particles will attract. As counterions in the solution tend to accumulate at the charged interface, they screen the ENP surface charge and effectively reduce the electrostatic repulsion between particles. The degree of charge screening is dependent on electrolyte concentration and valence of counterions. Charge neutralisation can also occur as a result of specific adsorption, when divalent and trivalent counterions or polyelectrolytes adsorb on the ENP surface within the Stern layer and reduce the effective surface charge. [29,35,43] Excess sorption of these counterions or polyelectrolytes in the Stern layer can result in charge reversal and particle restabilisation.^[29,35]

The classic Derjaguin–Landau–Verwey–Overbeek (DLVO) theory of colloidal stability is commonly invoked to describe the interfacial forces experienced between charged colloidal particles.[44,45] In essence, the theory states that the interaction energy between colloidal particles is the sum of van der Waals and electric double layer interactions. While van der Waals interactions are insensitive to changes in solution chemistry, the electric double layer interaction is strongly dependent on solution pH and ionic strength.^[35] As van der Waals and electric double layer interactions follow power-law and exponential-law relationships as a function of separation distance respectively, the magnitude of van der Waals interactions always exceeds electrostatic interactions at very small separation distances.^[29,35] For similarly charged particles and sufficiently low ionic strength conditions, the sum of these interaction energies results in the presence of an energy barrier that must be overcome by the kinetic energy of the particles before aggregation in the primary minimum can occur, as shown in Fig. 1. As the ionic strength increases, the energy barrier decreases until it is eliminated. At this point, the salt concentration has exceeded the critical coagulation concentration (CCC), resulting in favourable aggregation. The same logic applies in the case of deposition of colloids onto surfaces of larger solids. In this context, the pivotal salt concentration is known as the critical deposition concentration (CDC). At intermediate salt concentrations, an additional secondary minimum may be formed (Fig. 1).[46] Aggregation or deposition can take place through particle accumulation within the interfacial region of the secondary minimum (i.e. at this distance from the particle surface), but adhesion under these circumstances is much weaker than that which occurs when the two surfaces are sufficiently close to experience the energies of the primary minimum.^[35]

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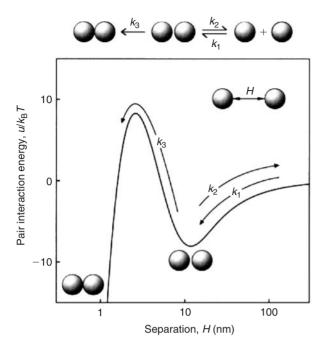


Fig. 1. Representative interaction energy, u, as a function of separation distance, H, between two similarly charged colloidal particles. The energy barrier is located at a separation distance of ~ 3 nm. $k_{\rm B}$ and T are the Boltzmann constant and absolute temperature respectively; k_1 , k_2 , and k_3 are rate constants for aggregation in the secondary minimum, redispersion after secondary aggregation, and aggregation in the primary minimum respectively. Reproduced from Behrens and Borkovec^[46] (copyright 2000, Elsevier BV).

Discrepancies between experimental results and predictions based on DLVO theory have been observed over the past few decades, particularly under conditions where repulsive electric double layer interactions predominate. [29,47–49] Under such conditions, the predicted aggregation and deposition kinetics (i.e. inverse stability ratios or attachment efficiencies) based on DLVO theory are frequently orders of magnitude lower than those obtained experimentally. Another discrepancy is that experiments have shown that colloidal stability is independent of particle size even when DLVO theory predicts that larger particles should be more stable than smaller ones. [47–49] These inconsistencies between experiments and theory have been attributed to surface roughness, surface charge heterogeneity, and non-DLVO forces such as hydrophobic interactions and structural forces. [29,47–49]

Steric stabilisation

Steric stabilisation refers to the enhancement in colloidal stability that occurs as a result of polymers adsorbed or grafted on the particle surface. As the particles approach each other, the interpenetration of the hydrophilic portion of the polymers that extends into the solution phase causes the displacement of water molecules into the bulk phase, resulting in an increase in free energy. [43] This repulsive interaction, which results from the polymer interpenetration, can be large enough to keep colloidal particles stable.

Steric stabilisation is the only mechanism at play for nonionic polymers,^[50] and the degree of stabilisation will be dependent on the surface density and the molecular weight of the polymers as well as the solubility of that portion of the polymer that extends into solution. As the extension of non-ionic polymers is largely insensitive to the presence of electrolytes, ^[43] the colloidal stability of particles coated with these polymers is not impacted by even large variations in salt concentrations. In cases where particles are only partially coated with polymers, bridging between particles (or flocculation) may occur instead. ^[51]

Conversely, when polyelectrolytes are adsorbed or grafted on the surface of particles, both steric and electrostatic repulsion could result to enhance the stability of particles. [52] The effect of both contributions is known as electrosteric stabilisation. Unlike steric stabilisation, the electrosteric effect is dependent on solution chemistry. The variation of solution pH, ionic strength, and divalent cation concentration can result in the conformational change of the surface-immobilised polyelectrolytes, as well as the overall surface charge of the coated particles. [53–55] Indeed, the challenge in modelling electrosteric repulsion is that both contributions are interdependent. [52,55] For instance, the degree of polyelectrolyte adsorption and polyelectrolyte conformation are dependent on the electric double layer, while the adsorption of polyelectrolytes itself can influence the double layer. [52]

In many cases, ENPs are provided (during synthesis or in subsequent modification) with surface coatings that enhance stability. For example, quantum dots can be coated with biocompatible polymers, [56,57] while zero-valent iron (ZVI) nanoparticles used for the treatment of groundwater contamination can be coated with synthetic or natural polymers to increase their mobility.^[58–60] Silver nanoparticles are often synthesised in the presence of polymeric capping agents in order to control their sizes. [61,62] In the natural environment, ENPs will also encounter NOM, and the effects of NOM on the stability of colloidal particles have been extensively investigated over the past few decades. [63–67] The nature of NOM varies significantly, however, as a result of the wide diversity of molecular weights and conformations, as well as from the negative polyelectrolytic nature of the humic and fulvic substances and polysaccharides that comprise a substantial fraction of NOM. Studies with these substances indicate that ENPs released into natural aquatic systems can adsorb NOM and that this can impart steric or electrosteric stabilisation. The situation becomes more complex, however, when NOM and ENPs also interact with other aqueous constituents, including multivalent cations and naturally occurring colloids.

Analytical and experimental methods

A comprehensive approach to understanding the environmental behaviour of ENPs is to correlate their physicochemical and electrokinetic properties with their colloidal stability. In the discussion that follows, we examine some prominent ENP characterisation techniques and highlight a few of the experimental methods used to measure colloidal stability. In this section, the discussion of particle imaging is treated in a cursory fashion, while techniques used to measure chemical composition, electrokinetic property, and aggregate structure are more thoroughly examined. Many but not all of the techniques presented here are used in the case studies of C_{60} nanoparticles and MWNTs discussed later in the review.

Characterisation techniques

Particle imaging

ENP morphology, size distribution, and dispersion state (i.e. primary particles, doublets, and higher-order aggregates) in solution may play important roles in controlling colloidal

behaviour. To characterise these attributes, techniques for direct visualisation, such as transmission electron microscopy (TEM), [68,69] scanning electron microscopy (SEM), [70,71] and atomic force microscopy (AFM)[72,73] can be used. TEM and SEM require ultra-high vacuum conditions and use electrons to render images (micrographs) of the ENPs being studied. AFM is performed under ambient or solution-phase conditions and renders images based on interactions between particles and a sharp tip. In all three techniques, ENP morphologies and size distributions may be assessed. In addition, qualitative appraisals of ENP dispersion states are also possible. Of the three techniques, TEM provides the highest magnification power and can be used to attain more detailed information on ENP structure. However, a drawback with all of these techniques is that sampling preparation methods and drying effects may change the aggregation state of the ENPs.

Elemental (bulk) composition

Energy dispersive X-ray spectroscopy (EDS)^[70] can be used to measure the elemental (bulk) composition of an entire ENP sample or a localised region, depending on the focus and magnification of the system.^[74] In the EDS process, an incident electron beam initiates the ejection of core-level electrons and the emission of element-dependent X-rays whose energy can be analysed to determine the ENPs' chemical composition. The analysis depth of EDS is $\sim 1.5 \,\mu m$ (although the exact depth will depend on the electron beam energy and the elements present in the particle) and the spot size is generally larger than 100 nm. It is important, however, to recognise the limitations of EDS. Given the analysis depth and spot size, data acquired on individual ENPs could also include signal from the substrate. [75] Additional techniques that can be used to determine the bulk composition of ENPs are inductively coupled plasma mass spectrometry (ICP-MS)^[76,77] and combustion analysis (CA).^[78] These techniques, which suffer similar problems of differentiation among particles, are also not especially useful for determining individual particle properties in heterogeneous materials.

Surface composition

X-ray photoelectron spectroscopy (XPS)^[79] is commonly used to characterise an ENP's surface composition. In this technique, a sample is irradiated with X-rays and the measured binding energies of the ejected photoelectrons provide information on the elements present at the surface. The surface specificity (3–10 nm) of this technique is a consequence of the extremely short inelastic mean free path of the ejected photoelectrons. For nanomaterials smaller than 10 nm, the surface specificity of XPS diminishes, and the compositional information obtained is comparable with the bulk measurements acquired with EDS. XPS for nanomaterials requires relatively large amounts of sample to sufficiently cover the entire analysis region (1 cm²) and reduce unwanted contributions from the substrate. As an internal check, XPS results should be consistent with reasonable expectations. For example, in the case of oxidised CNTs, the only elements detected should be oxygen and carbon. The detection of other elements is a strong indication that drying effects have led to the precipitation or adsorption of unwanted impurities on the sample surface.

In addition to surface composition, XPS can also be used to identify the nature and concentration of surface functional groups using either peak deconvolution methods or chemical derivatisation (CD) techniques. These two methods are described below.

- Peak deconvolution. Although the binding energy of photoelectrons ejected during X-ray irradiation is largely determined by the element and the core level from which they are ejected, their binding energies are also influenced (albeit to a lesser degree) by the element's local chemical environment. For example, photoelectrons ejected from carbon atoms bound to fluorine exhibit higher binding energies than photoelectrons ejected from hydrocarbons. In principle, this information can be used to deconvolute an XPS spectral envelope, such as the C(1s) region into its various components. Such interpretations can often be ambiguous, however, because of the close binding energy of different species present (e.g. C-O and C=O) on the surface, coupled with the limited resolution of X-ray photoelectron spectro meters. To overcome this problem, CD methods have been developed.
- 2. *Chemical derivatisation*. As an alternative to XPS peakfitting, CD techniques^[80,81] can be used to more accurately and precisely quantify specific functional groups present on surfaces. In CD, a targeted surface functional group reacts with a specific chemical (derivatising) reagent that contains a unique chemical tag that is not present on the ENP surface. The chemical tag is chosen so that its concentration can be quantified either by XPS or another analytical technique, such as fluorescence. When CD is used in conjunction with XPS, fluorine is often used as the chemical tag because it is not normally found in most ENPs and has a high X-ray photoelectron (XP) cross-section. By measuring the concentration of fluorine using XPS after a derivatising reaction, the concentration of the targeted functional groups can be determined. Fig. 2 shows the different derivatising reactions that have been used to titrate the concentration of hydroxyl, carbonyl, and carboxyl groups on oxidised CNTs. An analogous protocol has also been developed using 3,3,3-(trifluoropropyl)dimethylchlorosilane to assay the concentration of free hydroxyl groups on silica surfaces.^[82] Compared with XPS peak-fitting methods, CD offers a far more quantitative method to assay those functional groups for which selective derivatising reactions exist.

Surface charge

ENP surface charge $(C\,m^{-2})$ can be calculated by dividing an ENP's specific charge $(C\,g^{-1})$ by its specific surface area $(m^2\,g^{-1})$. Specific charge may be determined by titrating a known ENP mass with acid or base and solving a charge-balance equation. [83] An ENP's specific surface area can be estimated using the Brunauer–Emmett–Teller (BET) method, [84] although care should to be taken when interpreting BET measurements because the method is not believed to have absolute accuracy when pore sizes below 20 nm are being probed. [85] Also, the drying of ENP samples under vacuum conditions during BET measurements may cause the ENPs to aggregate in ways that prevent N₂ from entering occluded surfaces. This occlusion will result in an underestimation of the actual surface area available when ENPs are suspended in aqueous solutions. For these reasons, the interpretation of N₂ adsorption data with the BET approach is only suitable for providing relative comparison between ENP samples with full appreciation of its limitations. Care must also be taken in conducting and interpreting titration experiments with inorganic ENPs to account for possible particle dissolution, especially at low pH conditions. The advantage of potentiometric titration is that it provides a direct measurement

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Fig. 2. Surface oxides on carbon nanotubes (CNTs) that can be targeted by chemical derivatisation (CD), the derivatising agents, and the surface product of each derivatisation reaction. To determine the concentration of carboxyl groups on the CNTs' surface, both the nitrogen and fluorine concentrations determined from X-ray photoelectron spectroscopy (XPS) are needed.

of surface charge using readily available, cost-effective equipment. The technique does, however, require comparatively large quantities of ENPs $(0.1-2\,\mathrm{g\,L^{-1}})$. [86]

Electrokinetic properties

In electrokinetic techniques, an ENP's surface potential is approximated by its zeta (ζ) potential, as determined from some comparatively straightforward measures of electrophoretic mobility (EPM). Particle electrophoresis is a popular method used to determine a particle's EPM. This method involves the measurement of a particle's migration rate in an electric field of known strength, and for ENPs, this is most often measured using electrophoretic light-scattering systems. For spherical particles, EPM values may be converted to ζ potentials using well-known equations $^{[29]}$ or tabulated values. $^{[87]}$ In contrast to surface-charge titration, EPM measurements are conducted on dilute suspensions, so only small quantities of ENPs are needed. Other than electrophoresis, other options, such as electroacoustic $^{[88]}$ and agar-gel, $^{[89]}$ are also available.

Surface forces

Over the past two decades, particle–particle and particle–flat surface interactions have been investigated by performing force measurements using the AFM. [90] After attaching a colloidal particle on a tipless AFM cantilever, [91] the colloidal probe can be brought towards the substrate of interest and retracted from the substrate after contact occurs. By monitoring the deflection of the cantilever, the forces between the colloidal probe and substrate can be derived as a function of separation distance. As these measurements can be performed in aqueous media, they

allow the investigation of the interfacial interactions (including DLVO-type interactions and steric or electrosteric repulsion for polymer-coated surfaces) that determine the particle's aggregation and deposition behaviour. In principle, this method can also be used to probe ENP-ENP and ENP-solid-surface interactions. As the conventional method of attaching a colloidal particle to the AFM cantilever requires the manipulation of the particle under an optical microscope, a practical difficulty lies in the preparation of a nanometre-sized colloidal probe suitable for force measurements.^[92] However, some progress has been made in this regard, as demonstrated by studies that have used inorganic nanoparticles and CNTs attached to AFM cantilevers for force measurements as well as for imaging. [92–94] Alternatively, AFM force measurements between a colloidal probe whose composition is of interest and a 'lawn' of ENPs deposited on a flat substrate can be extrapolated to elucidate ENP-ENP and ENP-solid-surface interactions. [95]

Aggregate structure

When ENPs undergo homoaggregation in natural aquatic systems, the aggregate structure will influence their rate of sedimentation and transport behaviour. It is well known that the compactness of the aggregate structure is dependent on homoaggregation kinetics – diffusion-limited (favourable) aggregation results in open fractal structures, whereas reaction-limited (unfavourable) aggregation produces compact fractal structures. [96–98] Static light scattering (SLS) is a common technique used to study these aggregate structures. [99–102] It is a non-invasive technique that involves the irradiation of a suspension containing the aggregates with a laser beam and the

measurement of the intensities of the scattered light at various angles. The scattering intensities are plotted against the magnitude of the scattering vectors (which is a function of the scattering angle), and the slope obtained in the power-law region yields the fractal dimension, which can then be used as a metric to quantify the compactness of the aggregate structures. Typically, compact and open structures will yield fractal dimensions of ~2.1 and 1.8 respectively.^[97] In a recent study, Chen et al.^[103] determined the fractal dimensions of aggregates comprised of as-received and acid-treated single-walled CNTs to be 2.27 and 2.50 respectively by using SLS. Their results show that the shortened, acid-treated CNTs can form more compact aggregate structures compared with the longer, as-received CNTs. SLS can also be used to investigate the restructuring of aggregates that has been found to occur, as evident from the change in fractal dimensions with time. [99,102] For binary systems (i.e. systems comprising two types of colloidal particles), it is possible to obtain the fractal dimensions of heteroaggregates using SLS.[104,105] Care must be taken, however, to perform these measurements under solution conditions in which heteroaggregation occurs exclusively.

In addition to SLS, fractal dimensions of aggregates can be obtained by analysing the TEM or SEM images of aggregates. [101] However, this method involves the drying of the aggregates on a substrate, which is likely to result in the collapse of the aggregate structures. Also, a direct analysis of the TEM or SEM images would only allow the derivation of a two-dimensional fractal dimension, which may not be truly representative of the three-dimensional aggregate structure.

Measurements of aggregation and deposition kinetics Sedimentation properties

Sedimentation refers to the settling of particles and aggregates due to gravitation or forces of centrifugation. For a given settling force, the rate of sedimentation will depend on aggregate size, shape, and particle densities. Sedimentation experiments are frequently used to make qualitative, visual assessments of colloidal stability under different aquatic conditions. In conjunction with UV-visible spectroscopy or turbidity measurements, kinetic experiments may be conducted and compared among samples. [106–108] Through sedimentation experiments, for example, CCCs can be approximated by determining the minimum concentrations of electrolyte needed to remove a substantial amount of particles from the suspensions.[109,110] One drawback of the technique, however, is that quantitative comparisons with other studies cannot be made unless the particle concentrations and sedimentation conditions (e.g. time allowed for sedimentation to occur) are well controlled and carefully replicated.[111]

Homoaggregation kinetics

Time-resolved dynamic light scattering (TR-DLS) can measure temporal changes in the hydrodynamic diameter $(D_{\rm h})$ of aggregates during perikinetic homoaggregation. In these experiments, homoaggregation is typically induced by adding an electrolyte to the colloidal suspension being studied. As shown in Fig. 3, TR-DLS data can be used to construct aggregation profiles. In this profile, a linear increase in $D_{\rm h}$ with time is observed during the initial period of aggregation during which doublet formation is predominant. [54,112–114] However, during later stages of aggregation, the observed growth rate slows down owing to particle depletion.

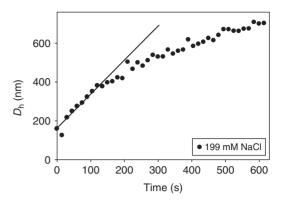


Fig. 3. Aggregation profile of oxidised multi-walled carbon nanotubes (MWNTs) (0.75 mg L⁻¹) in the presence of 199 mM NaCl at pH 6. Results of a linear fit are shown as a solid line between $D_h \sim 150$ and 400 nm. Reproduced from Smith et al.^[36] (copyright 2009, ACS Publications).

By fitting the initial linear region of Fig. 3, the initial particle growth rate, $(dD/dt)_{t\to 0}$, can be determined and used to obtain understanding about the aggregation rate constant, k, with Eqn 1:

$$k \propto \frac{1}{N_0} \left(\frac{dD_{\rm h}}{dt} \right)_{t \to 0} \tag{1}$$

where N_0 is the initial primary particle concentration.

For TR-DLS experiments that are conducted at the same initial particle concentration, Eqn 2 shows that $(dD_h/dt)_{t\to 0}$ values collected under different salt concentrations can be normalised to the diffusion-limited $(dD_h/dt)_{t\to 0}$ value $((dD_h/dt)_{t\to 0, fast})$ to yield an empirical inverse stability ratio, 1/W (or attachment efficiency, α):

$$\frac{1}{W} = \alpha = \frac{(dD_{\rm h}/dt)_{t\to 0}}{(dD_{\rm h}/dt)_{t\to 0, fast}} = \frac{k}{k_{fast}}$$
 (2)

where the subscript 'fast' refers to diffusion-limited (favourable) aggregation conditions.

When the inverse stability ratio is plotted as a function of electrolyte concentration, the result is an inverse stability profile, which may be fitted empirically to yield the CCC. [36,113] As the CCC represents the minimum amount of electrolyte needed to destabilise the suspension such that diffusion-limited aggregation occurs, it provides a useful metric of colloidal stability for ENPs.

Deposition kinetics

As ENPs can undergo heteroaggregation with other types of colloidal particles, it is of interest to measure and model the kinetics of heteroaggregation. Currently, a multiangle static and dynamic laser light-scattering technique exists for the determination of heteroaggregation rate constants from laboratory measurements. [30,31,33] A principal limitation, however, is that this technique requires a sophisticated light-scattering setup with multiple detectors that may not be commonly accessible in many laboratories.

In recent years, the quartz crystal microbalance (QCM) has emerged as an alternative method to measure the interactions of ENPs with other solid surfaces. The QCM comprises a flow cell that houses a quartz crystal. Because quartz is a piezoelectric material, the crystal can be excited to oscillate at its resonance and overtone frequencies under an applied electric potential difference. To investigate the interactions between ENPs and solid

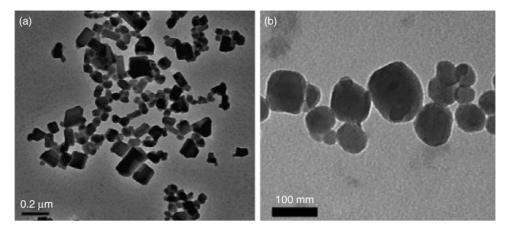


Fig. 4. Transmission electron microscopy (TEM) micrographs of C_{60} nanoparticles produced through solvent exchange with THF (a); and toluene (b). Reproduced from Fortner et al. [129] and Chen and Elimelech [113] (copyright 2005 and 2006, ACS Publications).

surfaces, an ENP suspension of interest is directed across the crystal surface under a constant flow rate. Depending on the solution chemistry, some ENPs will deposit on the crystal surface and cause a shift in the resonance and overtone frequencies. Within certain limits, the increase in total mass of the crystal due to ENP deposition is proportional to the shift in the resonance and overtone frequencies. [115] Hence, the deposition kinetics of the ENPs on the crystal surface can be derived from the rate of frequency shift.

Traditionally, the interactions between colloidal particles and other solid surfaces have been investigated through column filtration experiments. However, the interpretation of results from such systems is more complex because of the multiple processes affecting the retention of colloidal particles within the filter media. [116,117] These processes will depend not only on the surface-scale factors that influence DLVO interactions between particles and collectors, but also on physical properties such as the geometry and size of the particles and collectors and the hydrodynamics within the column. Usually, the flow chambers of the QCM use much simpler geometries, such as in radial stagnation point and parallel-plate flow systems. To date, the QCM has been used to study the deposition of fullerene C₆₀ nanoparticles, [113,118] titanium dioxide nanoparticles, [119] quantum dots, [120] viruses, [121] and ZVI nanoparticles [122,123] on silica surfaces.

Case studies

Fullerene C₆₀ nanoparticles

Buckminsterfullerene C_{60} exhibits unique physical, chemical, and electronic properties due to its spherical molecular structure. Since its discovery, [124] there has been growing interest in utilising this novel molecule for applications in diverse fields, including biomedical and environmental engineering. [19, 125] The major constraint to the use of fullerene C_{60} , however, is its extreme hydrophobicity, which limits its aqueous solubility. [126,127] Despite the relative insolubility of C_{60} molecules, the molecules are able to form colloidally stable C_{60} nanoparticles in aqueous solutions using well-established preparation methods. Because C_{60} is likely to take the form of nanoparticles when released into aqueous systems, the colloidal properties of these nanoparticles are expected to control their environmental fate and transport. As evidence from recent studies has shown that C_{60} nanoparticles

can be toxic, [128–131] a better understanding of their colloidal properties will also provide insights into nanoparticle—cell interactions, which may in turn help to elucidate the mechanisms of toxicity.

Preparation of C₆₀ colloids

In one of the earliest reports on the preparation of a C_{60} colloidal suspension, Scrivens et al.^[132] devised a synthesis method that involved first the dissolution of C₆₀ in benzene, followed by sequential dilution with tetrahydrofuran (THF) and acetone. The solvent-C₆₀ mixture was then introduced into water, which led to the formation of C₆₀ colloidal particles. The final suspension was distilled to remove most of the organic solvents. This method of 'solvent exchange' has since been used and modified by other researchers to synthesise C₆₀ nanoparticles. Other organic solvents that have been used include toluene^[54] and ethanol,^[133] and these solvents have been used either on their own or in various combinations. The type of solvents used and the parameters employed for the synthesis influenced the geometries and size distributions of the suspended C₆₀ nanoparticles. For instance, solvent-exchange using THF produces angular nanoparticles, whereas exchange using toluene produces nanoparticles that are more rounded (Fig. 4). The reasons for these differences in C_{60} nanoparticle structure are, however, not fully understood.

In more recent years, it has been shown that colloidally stable C_{60} nanoparticles can be produced by a 'top–down' approach that agitates bulk C_{60} crystals, breaking them into smaller C_{60} particles, some of which are expected to be in the nanometre-scale range. Some possible methods include the stirring of C_{60} crystals in water for a prolonged period of time (from 2 weeks to 5 months)[134–138] and ultrasonication of C_{60} crystals in water.[139,140]

Colloidal stability

The unusual colloidal stability of the supposedly hydrophobic C_{60} nanoparticles has been highlighted in numerous studies. [132,141–143] Specifically, C_{60} nanoparticles are found to be extremely stable to aggregation under low ionic-strength conditions regardless of the synthesis method used, and even in the absence of surfactants or stabilisers, they have been reported to remain colloidally stable for months. The electrokinetic properties of C_{60} nanoparticles produced using various methods have been investigated through EPM measurements. Interestingly,

these nanoparticles are found to exhibit negative EPMs under a wide range of pH conditions (from 2 to 12), [136,142–144] with the synthesis method playing an important role in determining the magnitude of the mobility. For instance, C₆₀ nanoparticles produced through extended stirring in water were more negatively charged than those produced through solvent-exchange methods using THF^[144] and toluene. [136] One possibility is that the difference in EPMs could be due to dissimilar surface chemistries resulting from the various preparation methods.

Influence of pH

More systematic electrokinetic studies have varied solution pH with the aim of understanding the source of the nanoparticle surface charge. C_{60} nanoparticles produced via either extended stirring in water or solvent exchange (with either THF or toluene) have exhibited increasingly negative EPM as pH is raised from 2 to 12 in the presence of monovalent electrolytes. [136,144] These results seem to imply that surface functional groups are a possible origin of C_{60} nanoparticle surface charge. It has also been suggested that the preferential adsorption of OH^- ions on C_{60} nanoparticle surfaces could potentially cause the nanoparticles to be more negatively charged under higher pH conditions owing to the greater availability of OH^- ions. [145] However, Brant et al. [144] measured the pH of the solution before and after the prolonged stirring required to disperse C_{60} in water and did not find any significant difference in the pH. At least for this system, it is therefore unlikely that the adsorption of OH^- ions is the origin of surface charge.

Influence of electrolytes

The electrokinetic properties of C₆₀ nanoparticles in different electrolytes have also been extensively investigated. In several studies, an increase in monovalent electrolyte concentration caused the EPM to become less negative, [136,144] consistent with charge screening effects. In most cases, divalent electrolytes were found to reduce the EPMs of C₆₀ nanoparticles more effectively compared with monovalent electrolytes, either owing to improved charge screening or possible specific adsorption of divalent cations. [136,144,146] However, in the presence of low concentrations of CaCl₂, Brant et al. [144] reported that charge reversal took place for C₆₀ nanoparticles produced through solvent exchange with THF. These C₆₀ nanoparticles, originally negatively charged at 0.01 mM CaCl₂, became positively charged at 0.1 mM CaCl₂, and subsequently became negatively charged again at ~0.3 mM CaCl₂. The authors attributed the charge reversal and re-reversal to the preferential adsorption of Ca²⁺ and Cl⁻ ions respectively. In a separate study, Wang et al.^[146] presented contradictory results, showing that nanoparticles produced by solvent exchange remained negatively charged over the same range of CaCl₂ concentrations studied by Brant et al. Further investigation is clearly required to verify the phenomenon of charge reversal (and re-reversal) in the presence of Ca^{2+} .

Aggregation kinetics

The first reported study on the aggregation behaviour of C_{60} nanoparticles was conducted by Mchedlov-Petrossyan et al. ^[142] The authors determined the aggregation kinetics of C_{60} nanoparticles synthesised through solvent exchange with toluene by monitoring the absorbance of aggregating C_{60} suspensions at 500 nm. At pH 5–6, the CCCs were found to be 85 mM NaCl and 4.1 mM $CaCl_2$. In a more recent study, TR-DLS was employed

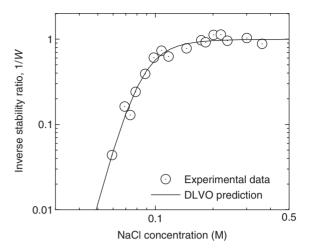


Fig. 5. Theoretical Derjaguin–Landau–Verwey–Overbeek (DLVO) prediction and experimentally derived inverse stability ratios of C₆₀ nanoparticles produced from solvent exchange with toluene as a function of NaCl concentration at pH 5.2. Aggregation experiments are carried out at 23°C. The critical coagulation concentration (CCC) is based on the intersection of the extrapolations of the reaction-limited and diffusion-limited regimes and estimated as 120 mM NaCl. Reproduced from Chen and Elimelech^[113] (copyright 2006, ACS Publications).

to investigate the aggregation kinetics of C₆₀ nanoparticles produced through a similar method over a range of NaCl and CaCl₂ concentrations at pH 5.2.^[113] Fig. 5 presents the inverse stability ratios (1/W) obtained as a function of NaCl concentration. The results from the latter study revealed reaction-limited (i.e. 1/W < 1) and diffusion-limited (i.e. 1/W = 1) aggregation in the presence of NaCl and CaCl₂, demonstrating that C₆₀ nanoparticles undergo classical aggregation behaviour typical of charged colloidal systems in aqueous solutions. The CCCs obtained (120 mM NaCl and 4.8 mM CaCl₂) were similar to, albeit slightly higher than the ones obtained by Mchedlov-Petrossyan et al. The dissimilarity in CCCs is likely due to slight variations in the preparation methods as well as the different experiments used to derive the aggregation kinetics. Through TR-DLS, C₆₀ nanoparticles produced by extended stirring in water were also shown to undergo classical aggregation behaviour.[136] However, C₆₀ nanoparticles produced by extended stirring were found to exhibit a much higher CCC (166 mM KCl) than C₆₀ nanoparticles produced through solvent exchange with toluene (40 mM KCl), both at pH 5.5. These results corroborate findings from EPM measurements that the colloidal stability of C₆₀ nanoparticles is influenced by the preparation method.

In their investigation of the aggregation kinetics of C₆₀ nanoparticles produced from solvent exchange with toluene as well as extended stirring in water, Chen and Elimelech^[113,136] compared the experimentally obtained inverse stability ratios with theoretical DLVO predictions. The authors found that particle stabilities for C₆₀ nanoparticles produced by either solvent exchange or water stirring were in good agreement with the DLVO predictions (Fig. 5), which is very uncommon in studies of aggregation and deposition kinetics of colloidal particles under conditions where repulsive electrostatic interactions predominate.^[29,47–49] Because DLVO theory is based on the simplifying assumption that the surface charge of the colloidal particle is uniformly distributed over the particle surface,^[29] discrepancies between experimental results and theoretical predictions have often been attributed to heterogeneities

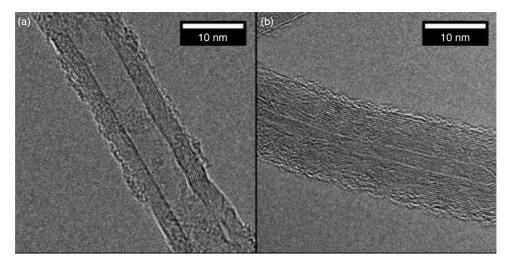


Fig. 6. Transmission electron microscopy (TEM) micrographs of a pristine multi-walled carbon nanotube (MWNT) (a); and an oxidised (3:1 v/v mixture of concentrated $\text{H}_2\text{SO}_4/\text{HNO}_3$) MWNT (b).

in surface charge. Hence, one possible reason for the good agreement between DLVO theory and experimental data in Chen and Elimelech's studies $^{[113,136]}$ could be that the charges are evenly distributed on the C_{60} nanoparticle surface.

Interactions with solid surfaces

The QCM has been used to investigate the deposition behaviour of C₆₀ nanoparticles prepared through solvent exchange with toluene. [118] The deposition kinetics of the nanoparticles on silica surfaces were found to increase with increasing NaCl and CaCl2 concentrations until transportlimited deposition occurred at the CDCs. At salt concentrations equal to or greater than the CDC value, the surface charges of the nanoparticles and solid surfaces are sufficiently screened such that favourable deposition occurs. The CDC was found to be 32.1 mM in NaCl (at pH 5.5), much higher than NaCl concentrations in freshwater systems, while the CDC in CaCl₂ was found to be 0.7 mM (at pH 5.5), which falls within the Ca²⁺ concentrations of hard freshwaters. Hence, the attachment of C₆₀ nanoparticles to silica surfaces is more likely to be controlled by the concentration of divalent cations in freshwater systems. These results from QCM experiments are in agreement with the ones from column filtration experiments that have also shown C₆₀ nanoparticle retention increasing with increasing salt concentrations.[146,147]

Influence of natural organic matter

In one of the earliest studies on the influence of NOM, Terashima and Nagao^[148] stirred C_{60} material in solutions containing humic and fulvic acids. They reported that C_{60} colloidal particles were produced in much higher concentrations in the presence of the organic matter and that the particles were smaller than the ones produced in pure water. [148] Similar findings on the reduction in C_{60} particle sizes in the presence of organic matter have also been observed in other studies. [137,149] The influence of humic acid was also found to be much greater than fulvic acid. [148] This observation was attributed to humic acid having a higher affinity with C_{60} because it has a lower charge density and a larger aromatic backbone compared with fulvic acid.

The influence of NOM on the colloidal stability of C_{60} nanoparticles was systematically investigated by Chen and Elimelech through TR-DLS. $^{[95]}$ A low concentration of humic acid

(1 mg L $^{-1}$ total organic carbon) was sufficient to significantly reduce the aggregation kinetics of C_{60} nanoparticles (at a concentration of 0.3 mg L $^{-1}$) in the presence of NaCl or MgCl $_2$ electrolytes. Even in a high ionic strength solution (650 mM NaCl), the inverse stability ratio obtained in the presence of humic acid was only \sim 0.1. Under such conditions, the charges on the humic acid-coated nanoparticles are likely to be effectively screened, indicating that the key stabilising mechanism is steric repulsion. However, in the presence of high concentrations of CaCl $_2$ electrolyte (>10 mM), bridging between the humic acid-coated C_{60} nanoparticles occurred, resulting in enhanced aggregation. These results clearly demonstrate that the influence of NOM on colloidal stability greatly depends on the solution chemistry.

Carbon nanotubes

CNTs are long, thin, hollow cylinders, composed of one (singlewalled CNTs, SWNTs) or many (MWNTs) concentric layers of graphenic carbon.^[4] Representative TEM images of MWNTs are presented in Fig. 6. Because of their unique electronic and mechanical properties, CNTs are being used in a growing number of commercial applications such as composite materials, [150] electronic devices, [151] and fuel cells. [152] For unmodified CNTs, surface hydrophobicity prohibits their dispersion in water, making them difficult to use in many applications. This behaviour of pristine CNTs is consistent with the relative insolubility of C₆₀ molecules. In contrast to C₆₀ nanoparticles, however, stable dispersions of CNTs are not created by stirring CNTs in water or conducting solvent exchange. Instead, their surfaces must be modified either by covalently attaching dissociable, hydrophilic functional groups (e.g. carboxylic acids, hydroxyls, amines, or amides) or by adsorbing stabilising macromolecules or surfactants. In the present discussion, we focus on the colloidal properties of MWNTs.

Surface oxidation

To create stable CNTs, the pristine nanomaterial is often treated with aggressive oxidants (e.g. HNO_3 , HNO_3/H_2SO_4 , $KMnO_4$, or H_2O_2), [153–156] cleaned, and dispersed using sonication. The principal effect of these oxidising treatments is to graft charged, hydrophilic functional groups (e.g. carboxyls and hydroxyls) into the exposed graphene sidewalls and at the tube

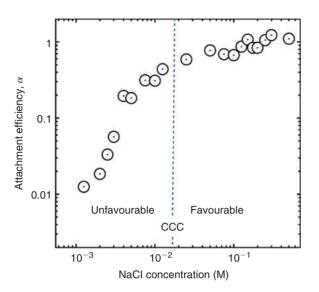


Fig. 7. Attachment efficiencies (or inverse stability ratios) of multi-walled carbon nanotubes (MWNTs) as a function of NaCl concentration at pH 6. Aggregation experiments are carried out at 23°C. The critical coagulation concentration (CCC) is based on the intersection of the extrapolations of the unfavourable (i.e. reaction-limited) and favourable (i.e. diffusion-limited) regimes and estimated as 25 mM NaCl. Reproduced from Saleh et al. [158] (copyright 2008, ACS Publications).

ends. In aqueous suspensions, these functional groups facilitate electrostatic stabilisation and favourable interactions with water molecules, thus promoting stability.

Influence of electrolytes

Both EPM measurements and surface-charge titrations have shown that acid treatments impart a negative charge to the CNT surface.^[86,157] Consistent with charge screening and charge neutralisation effects, increasing concentrations of monovalent and divalent electrolytes result in more rapid sedimentation of acid-washed MWNTs. [109] To shed additional light on the aggregation behaviour of MWNTs leading to sedimentation, Saleh et al. [158] and Smith et al. [36,86] have used TR-DLS to investigate MWNTs dispersed by high-power ultrasonication and acid treatment respectively. It is important to note that DLS measures particle size on the basis of a spherical particle approximation, far from the rod-like structure of CNTs. Although semi-flexible rod models have been developed for DLS that can provide information on MWNT length in non-aggregating systems, [159] a spherical approximation has been invoked to monitor the change in the particle size over time in the studies conducted on CNTs described here.

The results obtained using TR-DLS show that MWNTs exhibit reaction- and diffusion-limited aggregation regimes, comparable with trends observed in sedimentation studies (Fig. 7). [36,86,158] As stated, although DLVO theory can be applied in a quantitative way to spherical particles such as C_{60} nanoparticles (Fig. 5), its direct applicability to elongated particles and complex aggregates is less apparent and needs further investigation. However, results from studies on MWNT aggregation [36,86,158] show that qualitative tenets from DLVO theory do apply to electrolyte effects on the aggregation of MWNTs.

In the presence of NaCl, the CCC of acid-washed MWNTs (93 mM) is much higher than that of MWNTs prepared by high-power ultrasonication (25 mM) (Table 1). In similar solution

chemistries, the EPMs for the acid-washed and ultrasonicated MWNTs are -3.5×10^{-8} and -1.9×10^{-8} m 2 V $^{-1}$ s $^{-1}$ respectively. Thus, the larger absolute EPM values exhibited by the acid-washed MWNTs are consistent with their superior colloidal stability, which is believed to be a reflection of their higher surface concentration of dissociable functional groups.

Furthermore, analysis of Table 1 reveals that the acidwashed MWNTs appear to follow the Schulze–Hardy rule (SHR), which states that the CCC of highly charged particles is proportional to the counterion valence (Z) to the power of negative six (i.e. $CCC \propto Z^{-6}$). [29,43] Adherence to the SHR has also been observed in sedimentation studies with acid-washed CNTs. [109,110,160] In contrast, MWNTs prepared through high-power ultrasonication deviated from the SHR, yielding a $CCC \propto Z^{-3.3}$ relationship. This difference in the exponent on Z may be a reflection of the relatively low EPM exhibited by MWNTs prepared by ultrasonication, because the CCC is predicted to be proportional to Z^{-2} for particles with low ζ potentials. [29,43]

Influence of pH

The colloidal stability of acid-treated CNTs is sensitive to pH, and EPM studies have been conducted by several research groups [36,157,161–164] to determine the underlying cause. Typical trends for acid-treated CNTs are shown in Fig. 8. In this study, Schierz et al. [161] used MWNTs that were oxidised using a 3:1 H₂SO₄/HNO₃ mixture for 16 h (solid triangles) and 42 h (hollow triangles). As shown in the figure, ζ potentials increase in magnitude as pH is increased from 1 to 4. This is likely due to an increase in the number of charged carboxylate groups (–COO[–]) with increasing pH. Above pH 4, the ζ potential asymptotically levels out, although Saleh et al. [158] reported a steady increase in the magnitude of EPM of ultrasonicated MWNTs as the pH increased from 3 to 12.

Based on the many studies that have shown that the EPM of MWNTs remains constant above pH \sim 6, [36,157,163,164] the colloidal stability of MWNTs should be independent of pH in basic solutions. This hypothesis, however, contradicts visual observations from sedimentation studies that show decreases in the rates of MWNT aggregation and sedimentation with increasing pH. [36,165] Furthermore, Smith et al. [36] showed with TR-DLS that the CCCs of the acid-washed MWNTs increased linearly from 25 mM NaCl at pH 3 to 239 mM NaCl at pH 10. Saleh et al. [158] reported similar pH-dependent increases in MWNT stability using TR-DLS. Although variations in EPM with pH cannot explain the observed increases in MWNT colloidal stability under basic conditions, Smith et al.[36] have noted that the titrated surface charge increases systematically with increasing pH and correlates well with stability. The fundamental reasons why titrations provide a metric for colloidal stability while EPM measurements fail are unknown. Interestingly, a similar discrepancy between the variations in EPM and charge densities (calculated from potentiometric titration) with pH has been observed for colloidal silica.[166]

Influence of surface oxygen concentration

It is well known that oxygen-containing functional groups help stabilise CNTs in suspension. Indeed, qualitative assessments of colloidal stability due to oxidation have been made by conducting sedimentation studies. [163] Increased concentrations of protic oxygen functional groups (as measured by titration methods such as Boehm titrations [126,165]) have

Table 1. Critical coagulation concentrations (CCCs), exponents (n), and electrophoretic mobilities (EPMs) of multi-walled carbon nanotubes (MWNTs) prepared with two treatment methods

Treatment method	CCC ^A (mM)			<i>n</i> from $CCC \propto Z^n$ relationship		$EPM^{B} (10^{-8} m^{2} V^{-1} s^{-1})$	References
	Na ⁺	Mg^{2+}	Ca^{2+}	Mg^{2+}	Ca^{2+}		
Sonication ^C	25	1.5	2.6	-4.1	-3.3	-1.9	[158]
HNO ₃	93	1.8	1.2	-5.7	-6.3	-3.5	[36]

ACCC were derived from time-resolved dynamic light scattering (TR-DLS) experiments conducted at pH 5.8–6.0.

^CMWNTs were oxidised by high-power ultrasonication.

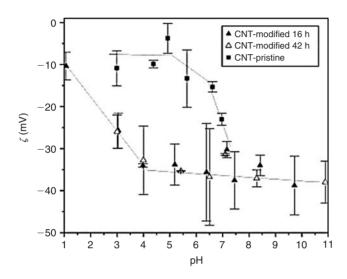


Fig. 8. Zeta (ζ) potentials of pristine and modified multi-walled carbon nanotubes (MWNTs) as a function of pH. All measurements were performed in the presence of 10 mM NaCl. The error bars represent two standard errors, obtained from three replicates. Filled triangles represent MWNTs oxidised in a 3:1 mixture of H_2SO_4 : HNO_3 for 16 h. Hollow triangles represent MWNTs oxidised in a 3:1 mixture of H_2SO_4 : HNO_3 for 42 h. Filled squares represent untreated MWNTs. Reproduced from Schierz et al. [161] (copyright 2009, Elsevier B.V.)

generally resulted in suspensions that were more resistant to electrolyte-induced aggregation. [165]

More quantitative structure–property relationships linking surface chemistry to colloidal behaviour have recently been developed by Smith et al. [86] using a suite of oxidised MWNTs. In this study, XPS was used to quantify the surface oxygen concentration and determine the distribution of carboxyl, hydroxyl, and carbonyl groups on the MWNT surface. As shown in Fig. 9, the CCC increased linearly with MWNTs' surface oxygen concentration at three different pH values. Furthermore, Smith et al. [86] showed that negatively charged surface carboxyl groups played the dominant role in stabilising CNT suspensions, although the other oxygen-containing functionalities were also found to contribute to stability.

Influence of natural organic matter

Recent investigations have shown that the colloidal stability of pristine CNTs is significantly enhanced by the adsorption of NOM. Indeed, Hyung et al. [167] reported that adsorbed NOM stabilised MWNTs more effectively than the surfactant sodium dodecyl sulfate (SDS). Similar to previously discussed results from NOM-C₆₀ studies, [148] results from studies in NOM-MWNT systems showed that colloidal suspensions of

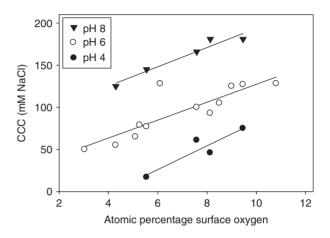


Fig. 9. Influence of surface oxygen concentration on the critical coagulation concentration (CCC) of oxidised multi-walled carbon nanotubes (MWNTs) at pH 4, 6, and 8. Reproduced from Smith et al.^[86] (copyright 2009, ACS Publications).

individually dispersed MWNTs could be produced by mere agitation (stirring and shaking). Similar results have been noted using a tannic acid surrogate for NOM.^[168] These observations are in marked contrast to the case of oxidised MWNTs where aggressive sonication methods were required for dispersion in the absence of NOM.

To develop a mechanistic understanding of the effect that NOM has on MWNT stability, the sorption of humic and fulvic acids with MWNTs has been studied.[169-171] Results from these studies reveal that MWNT sorption capacity for humics is greater than that for fulvics, [169] similarly to the findings of Terashima and Nagao^[148] in their study of NOM-C₆₀ suspensions. The stronger affinity observed for humics was attributed to their greater aromatic character, which resulted in enhanced π - π interactions with the MWNTs' surface. [169,170] The adsorption of fulvics was also shown to increase with increasing ionic strength and decrease with increasing pH.[171] Using TR-DLS, Saleh et al. [158] were able to compare the inverse stability ratios of MWNTs in the presence and absence of adsorbed humic acid. With the humic coating, the inverse stability ratios were one to two orders of magnitude lower than those observed for uncoated MWNTs, indicating a significant increase in colloidal stability. Despite the differences in colloidal behaviour, EPM measurements yielded similar results for both samples. This observation suggested that the stabilisation effect of NOM is purely a result of steric stabilisation.

In general, owing to the dynamic nature of sorption under various solution-phase conditions and the fact that fulvics and humics adsorb to different extents, predicting the stability of NOM-coated CNTs remains a challenge. However, Hyung

^BEPM measurements were conducted in the presence of 10 mM NaCl at pH 6–7.

et al. [169] have suggested that the stability conferred to the CNTs by NOM depends primarily on the concentration of NOM and not its exact composition. Clearly, additional studies are needed to uncover the detailed interactions of different NOM fractions with CNTs.

Comparing origins of surface charge of C_{60} nanoparticles and CNTs

Much more is known about the origin of surface charge for CNTs than for C₆₀ nanoparticles. Studies that have systematically varied the degree of CNT surface oxidation have verified surface functional groups to be an important cause of surface charge for these materials. Various surface-charging mechanisms for C_{60} nanoparticles have been hypothesised and discussed, [136,143–145] and one clue is found in the apparently uniform charge distribution, as evidenced by the consistency of aggregation behaviour with DLVO theory. [113,136] Some of the suggested mechanisms include the presence of dissociable surface functional groups, the preferential adsorption of OH- or other anions, and charge transfer from solvent molecules. It is important to realise that there could be more than one origin of surface charge and that the contribution from each origin could be different for C₆₀ nanoparticles synthesised through different methods. Experimental attempts to identify the origins of the surface charge on C₆₀ nanoparticles have recently been made. For example, the EPMs of C₆₀ nanoparticles synthesised through solvent exchange and extended stirring in water were measured and compared in solutions containing different anions (including Cl^{-} , I^{-} , and SO_4^{2-}). [136,144] The EPMs were not significantly different in the various electrolytes, [136] and no characteristic maximum (an indication of preferential adsorption^[172]) was observed when EPMs were measured as a function of electrolyte concentration.[144] Thus, it seems unlikely that anion adsorption is the origin of the surface charge. Based on the results obtained by Brant et al., [144] who observed no significant change in pH after the mixing process, the adsorption of OH- ions has also been ruled out as a possible charging mechanism for C₆₀ nanoparticles formed from extended stirring in water.

Prior studies have shown that oxidative treatments can result in the formation of oxygen-containing functional groups on CNTs. Because the degree of curvature for CNTs is confined to only a single dimension, C₆₀ is expected to be more susceptible to chemical modification than CNTs.[19] In a recent study, Chen and Elimelech^[136] used XPS analysis to reveal an increase in oxygen content on the surface of C₆₀ material that had been stirred in water for a prolonged period of time. In another study, Labille et al. [140] reported the presence of hydroxyl functional groups on C₆₀ nanoparticles produced through sonication in water based on their results from Fourier transform infrared (FTIR) and solid-state ¹H nuclear magnetic resonance (NMR) analyses. In addition, numerous electrokinetic studies have shown that C_{60} nanoparticles become more negatively charged at higher pH conditions.^[136,144,145] Thus, all evidence seems to point to the fact that ionic surface functional groups are at least in part responsible for the surface charge on C₆₀ nanoparticles. However, it is still unclear why the nanoparticles exhibit negative charge even at pH 2 when most functional groups should remain protonated. [135,136,144]

Clearly, detailed and quantitative knowledge of the surface chemistry of C_{60} nanoparticles is a necessary step to reveal the origin of the surface charge on these ENPs. However, to date, no studies have been conducted to provide functional group-specific

information on the surface chemistry of C_{60} nanoparticles. In this regard, XPS performed in conjunction with CD would be a potential method suitable to test for and identify the nature and concentration of surface functionalities on C_{60} nanoparticles. The key challenge is to produce sufficient amounts of dried C_{60} nanoparticles (at least a few milligrammes for XPS analysis) especially when the current synthesis techniques only yield C_{60} nanoparticle suspensions with concentrations ranging from a few to tens of milligrams per litre.

Research needs and future directions

It is clear from the discussion in this review article that one necessary requirement to rationalise the colloidal stability of ENPs is that the physicochemical properties of the ENPs themselves must be rigorously characterised and understood. In particular, there is a need for a better understanding of the surface chemistry of ENPs, which is known to depend on the synthesis and preparation techniques employed as well as subsequent modifications. In addition to deliberate surface modification, ENPs released into natural and engineered systems may undergo light-based transformations, oxidations or interactions with other aquatic species that can result in changes to their surface composition. [173–175] In this review, we have shown that XPS used in conjunction with CD can be a powerful technique for the determination of the surface functionalities of ENPs. The ability to accurately identify and quantify the surface functional groups appears to open the door to develop structure-property relationships that can be used to predict the colloidal stability as well as other properties (e.g. toxicity and reactivity) of ENPs based on their surface

Currently, most studies have focussed on the homoaggregation kinetics of ENPs. However, heteroaggregation between ENPs and naturally occurring colloids is almost certain to be a major process in natural aquatic systems. To date, quantitative studies on the heteroaggregation kinetics of ENPs are limited owing to the complexity of the systems involved. QCM has recently emerged as an option that can allow the direct measurements of ENP deposition kinetics on solid surfaces, but this approach still lacks the desired ability to capture the heteroaggregation kinetics between ENP and colloidal particles of varying types and sizes. Thus, there is a need for the development of novel and more accessible experimental techniques for the study of heteroaggregation between ENPs and other colloidal particles.

The homoaggregation and heteroaggregation (deposition) behaviour of ENPs are controlled by ENP-ENP and ENP-solid-surface interactions respectively. Even though current experimental results have shown that the aggregation and deposition behaviour of ENPs is qualitatively in agreement with DLVO theory, more study on the effects of non-DLVO forces is required. The most direct approach is to probe the interfacial forces between ENPs (or between ENPs and solid surfaces) under varying solution conditions. Recent developments have allowed the use of AFM force measurements between ENPs and solid surfaces. However, the preparation of a nanometre-sized colloidal probe for such measurements still remains a complicated task and an as yet unsolved challenge for many materials.

The mobility, bioavailability, and toxicity of ENPs are controlled by the aggregate structure in which they reside. Even though the influence of aggregation kinetics on the compactness of aggregate structures formed from monodisperse spherical particles is well known, few studies have been conducted on the

aggregate structures formed from suspensions of polydisperse and non-spherical particles, which are arguably the more typical cases for ENP use and release. For instance, it is expected that the bundling of CNTs will result in a more compact structure than the random cross-linking between them. Similar questions can be asked about the heteroaggregate structures formed from particles of different shapes and sizes, such as the possible combination of CNTs and naturally occurring silica nanoparticles. Cryogenic TEM (Cryo-TEM) offers the potential to acquire detailed molecular-scale images of ENP aggregation processes as well as the aggregates formed in situ under aqueous conditions. [176–178]

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References

- L. C. Qin, X. L. Zhao, K. Hirahara, Y. Miyamoto, Y. Ando, S. Iijima, Materials science – the smallest carbon nanotube. *Nature* 2000, 408, 50. doi:10.1038/35040699
- [2] T. R. Society, Nanoscience and Nanotechnologies: Opportunities and Uncertainties 2005 (The Royal Society: London).
- [3] K. R. Brown, D. G. Walter, M. J. Natan, Seeding of colloidal Au nanoparticle solutions. 2. Improved control of particle size and shape. *Chem. Mater.* 2000, 12, 306. doi:10.1021/CM980065P
- [4] S. Iijima, Helical microtubules of graphitic carbon. *Nature* 1991, 354, 56. doi:10.1038/354056A0
- [5] J. Shen, Y. Hu, M. Shi, X. Lu, C. Qin, C. Li, M. Ye, Fast and facile preparation of graphene oxide and reduced graphene oxide nanoplatelets. *Chem. Mater.* 2009, 21, 3514. doi:10.1021/ CM901247T
- [6] P. Short, M. McCoy, Companies invest in nanotubes. Chem. Eng. News 2007, 85, 20.
- J. Oliver, A. Crull, Nanoparticle News Review, 2008 2009
 (BCC Research). Available at http://www.bccresearch.com/report/ NAN004J.html [Verified 22 January 2010]
- [8] A. McWilliams, Nanotechnology: a Realistic Market Assessment 2004 (BCC Research). Available at http://www.bccresearch. com/report/NAN031C.html [Verified 22 January 2010]
- [9] The Project on Emerging Nanotechnologies (Woodrow Wilson International Center for Scholars and the Pew Charitable Trusts). Available at http://www.nanotechproject.org/inventories/consumer/ analysis_draft/ [Verified 22 January 2010]
- [10] J. Theron, J. A. Walker, T. E. Cloete, Nanotechnology and water treatment: applications and emerging opportunities. *Crit. Rev. Microbiol.* 2008, 34, 43. doi:10.1080/10408410701710442
- [11] M. Otto, M. Floyd, S. Bajpai, Nanotechnology for site remediation. Remediation 2008, 19, 99. doi:10.1002/REM.20194
- [12] S. N. Varadhi, H. Gill, L. J. Apoldo, K. Liao, R. A. Blackman, W. K. Wittman, Full-scale nanoiron injection for treatment of groundwater contaminated with chlorinated hydrocarbons, in *Natural Gas Technologies 2005 Conference, Orlando, FL* 2005 (Pars Environmental Inc.: Orlando, FL).
- [13] P. G. Tratnyek, R. L. Johnson, Nanotechnologies for environmental cleanup. *Nano Today* **2006**, *1*, 44. doi:10.1016/S1748-0132(06) 70048-2
- [14] B. Karn, T. Kuiken, M. Otto, Nanotechnology and in situ remediation: a review of the benefits and potential risks. *Environ. Health Perspect.* 2009, 117. doi:10.1289/EHP.0900793

- [15] B. Nowack, Pollution prevention and treatment using nanotechnology, in *Nanotechnology, Volume 2: Environmental Aspects* (Ed. H. Krug) 2008 (Wiley-VCH: Weinheim, Germany).
- [16] R. A. Freitas, What is nanomedicine? Nanomedicine 2005, 1, 2.
- [17] L. Lacerda, A. Bianco, M. Prato, K. Kostarelos, Carbon nanotubes as nanomedicines: from toxicology to pharmacology. Adv. Drug Deliv. Rev. 2006, 58, 1460. doi:10.1016/J.ADDR.2006.09.015
- [18] Office of Solid Waste and Emergency Response, Selected Sites Using or Testing Nanoparticles for Remediation 2008 (US EPA). Available at http://clu-in.org/download/remed/nano-site-list.pdf [Verified 22 January 2010]
- [19] M. S. Mauter, M. Elimelech, Environmental applications of carbon-based nanomaterials. *Environ. Sci. Technol.* 2008, 42, 5843. doi:10.1021/ES8006904
- [20] L. Li, Y. Xing, Pt-Ru nanoparticles supported on carbon nanotubes as methanol fuel cell catalysts. J. Phys. Chem. C 2007, 111, 2803. doi:10.1021/JP0655470
- [21] Y. Xing, Synthesis and electrochemical characterization of uniformly dispersed high-loading Pt nanoparticles on sonochemically treated carbon nanotubes. J. Phys. Chem. B 2004, 108, 19255. doi:10.1021/JP046697I
- [22] A. R. Köhler, C. Som, A. Helland, F. Gottschalk, Studying the potential release of carbon nanotubes throughout the application life cycle. *J. Clean. Prod.* 2008, 16, 927. doi:10.1016/J.JCLEPRO. 2007.04.007
- [23] N. C. Mueller, B. Nowack, Exposure modeling of engineered nanoparticles in the environment. *Environ. Sci. Technol.* 2008, 42, 4447. doi:10.1021/ES7029637
- [24] X. Zhu, L. Zhu, Z. Duan, R. Qi, Y. Li, Y. Lang, Comparative toxicity of several metal oxide nanoparticle aqueous suspensions to zebrafish (*Danio rerio*) early developmental stage. *J. Environ. Sci. Health Part A Tox. Hazard. Subst. Environ. Eng.* 2008, 43, 278. doi:10.1080/10934520701792779
- [25] S. Kang, M. S. Mauter, M. Elimelech, Microbial cytotoxicity of carbon-based nanomaterials: implications for river water and wastewater effluent. *Environ. Sci. Technol.* 2009, 43, 2648. doi:10.1021/ES8031506
- [26] S. Kang, M. S. Mauter, M. Elimelech, Physicochemical determinants of multiwalled carbon nanotube bacterial cytotoxicity. Environ. Sci. Technol. 2008, 42, 7528. doi:10.1021/ES8010173
- [27] B. J. Panessa-Warren, M. M. Maye, J. B. Warren, K. M. Crosson, Single-walled carbon nanotube reactivity and cytotoxicity following extended aqueous exposure. *Environ. Pollut.* 2009, 157, 1140. doi:10.1016/J.ENVPOL.2008.12.028
- [28] T. M. Sager, D. W. Porter, V. A. Robinson, W. G. Lindsley, D. E. Schwegler-Berry, V. Castranova, Improved method to disperse nanoparticles for in vitro and in vivo investigation of toxicity. *Nanotoxicology* 2007, 1, 118. doi:10.1080/17435390701381596
- [29] M. Elimelech, J. Gregory, X. Jia, R. A. Williams, Particle Deposition and Aggregation: Measurement, Modelling and Simulation 1995 (Butterworth-Heinemann: Oxford, UK).
- [30] W. L. Yu, M. Borkovec, Distinguishing heteroaggregation from homoaggregation in mixed binary particle suspensions by multiangle static and dynamic light scattering. J. Phys. Chem. B 2002, 106, 13106. doi:10.1021/JP021792H
- [31] W. Lin, M. Kobayashi, M. Skarba, C. D. Nu, P. Galletto, M. Borkovec, Heteroaggregation in binary mixtures of oppositely charged colloidal particles. *Langmuir* 2006, 22, 1038. doi:10.1021/ LA0522808
- [32] P. Galletto, W. Lin, M. Borkovec, Measurement of heteroaggregation rate constants by simultaneous static and dynamic light scattering. *Phys. Chem. Chem. Phys.* 2005, 7, 1464. doi:10.1039/ B417761D
- [33] W. L. Yu, E. Matijević, M. Borkovec, Absolute heteroaggregation rate constants by multiangle static and dynamic light scattering. *Langmuir* 2002, 18, 7853. doi:10.1021/LA0203382
- [34] K. M. Yao, M. M. Habibian, C. R. O'Melia, Water and waste water filtration – concepts and applications. *Environ. Sci. Technol.* 1971, 5, 1105. doi:10.1021/ES60058A005

- [35] J. Israelachvili, Intermolecular and Surface Forces 1991 (Academic Press: London, UK).
- [36] B. Smith, K. Wepasnick, K. E. Schrote, A. H. Bertele, W. P. Ball, C. O'Melia, D. H. Fairbrother, Colloidal properties of aqueous suspensions of acid-treated, multi-walled carbon nanotubes. *Environ. Sci. Technol.* 2009, 43, 819. doi:10.1021/ES802011E
- [37] D. Fornasiero, F. Grieser, The kinetics of electrolyte-induced aggregation of Carey Lea silver colloids. *J. Colloid Interface Sci.* 1991, 141, 168. doi:10.1016/0021-9797(91)90312-V
- [38] Z. S. Pillai, P. V. Kamat, What factors control the size and shape of silver nanoparticles in the citrate ion reduction method? *J. Phys. Chem. B* 2004, 108, 945. doi:10.1021/JP037018R
- [39] Y. Rong, H. Z. Chen, G. Wu, M. Wang, Preparation and characterization of titanium dioxide nanoparticle/polystyrene composites via radical polymerization. *Mater. Chem. Phys.* 2005, 91, 370. doi:10.1016/J.MATCHEMPHYS.2004.11.042
- [40] K. Yang, B. S. Xing, Sorption of phenanthrene by humic acidcoated nanosized TiO₂ and ZnO. *Environ. Sci. Technol.* 2009, 43, 1845. doi:10.1021/ES802880M
- [41] L. K. Limbach, Y. C. Li, R. N. Grass, T. J. Brunner, M. A. Hintermann, M. Muller, D. Gunther, W. J. Stark, Oxide nanoparticle uptake in human lung fibroblasts: effects of particle size, agglomeration, and diffusion at low concentrations. *Environ. Sci. Technol.* 2005, 39, 9370. doi:10.1021/ES051043O
- [42] A. K. Gupta, M. Gupta, Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. *Biomaterials* 2005, 26, 3995. doi:10.1016/J.BIOMATERIALS.2004.10.012
- [43] R. J. Hunter, Foundations of Colloid Science 2002 (Oxford University Press: Oxford, UK).
- [44] B. V. Derjaguin, L. D. Landau, Theory of the stability of strongly charged lyophobic sols and of the adhesion of strongly charged particles in solutions of electrolytes. *Acta Physicochim. URSS* 1941, 14, 733.
- [45] E. J. W. Verwey, J. T. G. Overbeek, Theory of the Stability of Lyophobic Colloids 1948 (Elsevier: Amsterdam).
- [46] S. H. Behrens, M. Borkovec, Influence of the secondary interaction energy minimum on the early stages of colloidal aggregation. J. Colloid Interface Sci. 2000, 225, 460. doi:10.1006/JCIS. 2000.6780
- [47] M. Elimelech, C. R. O'Melia, Effect of particle-size on collision efficiency in the deposition of Brownian particles with electrostatic energy barriers. *Langmuir* 1990, 6, 1153. doi:10.1021/ LA00096A023
- [48] S. H. Behrens, M. Borkovec, P. Schurtenberger, Aggregation in charge-stabilized colloidal suspensions revisited. *Langmuir* 1998, 14, 1951. doi:10.1021/LA971237K
- [49] S. H. Behrens, D. I. Christl, R. Emmerzael, P. Schurtenberger, M. Borkovec, Charging and aggregation properties of carboxyl latex particles: experiments versus DLVO theory. *Langmuir* 2000, 16, 2566. doi:10.1021/LA991154Z
- [50] D. H. Napper, Steric stabilization. J. Colloid Interface Sci. 1977, 58, 390. doi:10.1016/0021-9797(77)90150-3
- [51] E. Dickinson, L. Eriksson, Particle flocculation by adsorbing polymers. Adv. Colloid Interface Sci. 1991, 34, 1. doi:10.1016/0001-8686(91)80045-L
- [52] M. B. Einarson, J. C. Berg, Electrosteric stabilization of colloidal latex dispersions. J. Colloid Interface Sci. 1993, 155, 165. doi:10.1006/JCIS.1993.1022
- [53] A. Pettersson, G. Marino, A. Pursiheimo, J. B. Rosenholm, Electrosteric stabilization of Al₂O₃, ZrO₂, and 3Y-ZrO₂ suspensions: effect of dissociation and type of polyelectrolyte. *J. Colloid Interface Sci.* 2000, 228, 73. doi:10.1006/JCIS.2000.6939
- [54] K. L. Chen, S. E. Mylon, M. Elimelech, Aggregation kinetics of alginate-coated hematite nanoparticles in monovalent and divalent electrolytes. *Environ. Sci. Technol.* 2006, 40, 1516. doi:10.1021/ ES0518068
- [55] G. Fritz, V. Schadler, N. Willenbacher, N. J. Wagner, Electrosteric stabilization of colloidal dispersions. *Langmuir* 2002, 18, 6381. doi:10.1021/LA015734J

- [56] I. Yildiz, B. McCaughan, S. F. Cruickshank, J. F. Callan, F. M. Raymo, Biocompatible CdSe-ZnS core-shell quantum dots coated with hydrophilic polythiols. *Langmuir* 2009, 25, 7090. doi:10.1021/LA900148M
- [57] W. W. Yu, E. Chang, J. C. Falkner, J. Y. Zhang, A. M. Al-Somali, C. M. Sayes, J. Johns, R. Drezek, V. L. Colvin, Forming biocompatible and non-aggregated nanocrystals in water using amphiphilic polymers. J. Am. Chem. Soc. 2007, 129, 2871. doi:10.1021/JA067184N
- [58] T. Phenrat, N. Saleh, K. Sirk, H. J. Kim, R. D. Tilton, G. V. Lowry, Stabilization of aqueous nanoscale zero-valent iron dispersions by anionic polyelectrolytes: adsorbed anionic polyelectrolyte layer properties and their effect on aggregation and sedimentation. J. Nanopart. Res. 2008, 10, 795. doi:10.1007/S11051-007-9315-6
- [59] A. Tiraferri, K. L. Chen, R. Sethi, M. Elimelech, Reduced aggregation and sedimentation of zero-valent iron nanoparticles in the presence of guar gum. *J. Colloid Interface Sci.* 2008, 324, 71. doi:10.1016/J.JCIS.2008.04.064
- [60] S. R. Kanel, R. R. Goswami, T. P. Clement, M. O. Barnett, D. Zhao, Two-dimensional transport characteristics of surface stabilized zero-valent iron nanoparticles in porous media. *Environ. Sci. Technol.* 2008, 42, 896. doi:10.1021/ES071774J
- [61] H. H. Huang, X. P. Ni, G. L. Loy, C. H. Chew, K. L. Tan, F. C. Loh, J. F. Deng, G. Q. Xu, Photochemical formation of silver nanoparticles in poly(N-vinylpyrrolidone). *Langmuir* 1996, 12, 909. doi:10.1021/LA950435D
- [62] L. Quaroni, G. Chumanov, Preparation of polymer-coated functionalized silver nanoparticles. J. Am. Chem. Soc. 1999, 121, 10642. doi:10.1021/JA992088O
- [63] C. L. Tiller, C. R. O'Melia, Natural organic-matter and colloidal stability – models and measurements. *Colloids Surf. A Physicochem. Eng. Asp.* 1993, 73, 89. doi:10.1016/0927-7757(93)80009-4
- [64] E. Tipping, D. C. Higgins, The effect of adsorbed humic substances on the colloid stability of hematite particles. *Colloids Surf.* 1982, 5, 85. doi:10.1016/0166-6622(82)80064-4
- [65] R. Amal, J. A. Raper, T. D. Waite, Effect of fulvic acid adsorption on the aggregation kinetics and structure of hematite particles. *J. Colloid Interface Sci.* 1992, 151, 244. doi:10.1016/ 0021-9797(92)90255-K
- [66] I. Heidmann, I. Christl, R. Kretzschmar, Aggregation kinetics of kaolinite-fulvic acid colloids as affected by the sorption of Cu and Pb. Environ. Sci. Technol. 2005, 39, 807. doi:10.1021/ ES049387M
- [67] J. Buffle, K. J. Wilkinson, S. Stoll, M. Filella, J. W. Zhang, A generalized description of aquatic colloidal interactions: the threecolloidal component approach. *Environ. Sci. Technol.* 1998, 32, 2887. doi:10.1021/ES980217H
- [68] D. B. Williams, C. B. Carter, Transmission Electron Microscopy: a Textbook for Materials Science, 1st edn 2004 (Springer: New York).
- [69] G. G. Leppard, Nanoparticles in the environment as revealed by transmission electron microscopy: detection, characterisation and activities. *Current Nanoscience* 2008, 4, 278. doi:10.2174/ 157341308785161109
- [70] J. Goldstein, D. E. Newbury, D. C. Joy, C. E. Lyman, P. Echlin, E. Lifshin, L. C. Sawyer, J. R. Michael, Scanning Electron Microscopy and X-ray Microanalysis, 3rd edn 2003 (Springer: New York).
- [71] Y. Sun, Y. Xia, Shape-controlled synthesis of gold and silver nanoparticles. *Science* 2002, 298, 2176. doi:10.1126/SCIENCE. 1077229
- [72] J. R. Lead, K. J. Wilkinson, Aquatic colloids and nanoparticles: current knowledge and future trends. *Environ. Chem.* 2006, 3, 159. doi:10.1071/EN06025
- [73] C. N. R. Rao, K. Biswas, Characterization of nanomaterials by physical methods. *Annu. Rev. Anal. Chem.* 2009, 2, 435.
- [74] H. Yang, P. H. Holloway, Enhanced photoluminescence from CdS: Mn/ZnS core/shell quantum dots. Appl. Phys. Lett. 2003, 82, 1965. doi:10.1063/1.1563305

- [75] H. W. Kim, S. H. Shim, Branched structures of tin oxide onedimensional nanomaterials. *Vacuum* 2008, 82, 1395. doi:10.1016/ J.VACUUM.2008.03.074
- [76] H. J. Lee, S. Y. Yeo, S. H. Jeong, Antibacterial effect of nanosized silver colloidal solution on textile fabrics. *J. Mater. Sci.* 2003, 38, 2199. doi:10.1023/A:1023736416361
- [77] R. H. Yang, L. W. Chang, J. P. Wu, M. H. Tsai, H. J. Wang, Y. C. Kuo, T. K. Yeh, C. S. Yang, P. Lin, Persistent tissue kinetics and redistribution of nanoparticles, Quantum Dot 705, in mice: ICP-MS quantitative assessment. *Environ. Health Perspect.* 2007, 115, 1339.
- [78] C. Baleizão, B. Gigante, H. Garcia, A. Corma, Vanadyl salen complexes covalently anchored to single-wall carbon nanotubes as heterogeneous catalysts for the cyanosilylation of aldehydes. *J. Catal.* 2004, 221, 77. doi:10.1016/J.JCAT.2003.08.016
- [79] J. C. Vickerman, I. S. Gilmore (Eds), Surface analysis, in *The Principal Techniques*, 2nd edn 2009 (Wiley: Chichester, UK).
- [80] L. A. Langley, D. H. Fairbrother, Effect of wet chemical treatments on the distribution of surface oxides on carbonaceous materials. *Carbon* 2007, 45, 47. doi:10.1016/J.CARBON.2006.08.008
- [81] L. A. Langley, D. E. Villanueva, D. H. Fairbrother, Quantification of surface oxides on carbonaceous materials. *Chem. Mater.* 2006, 18, 169. doi:10.1021/CM051462K
- [82] N. M. Washton, S. L. Brantley, K. T. Mueller, Probing the molecular-level control of aluminosilicate dissolution: a sensitive solid-state NOM proxy for reactive surface area. *Geochim. Cosmochim. Acta* 2008, 72, 5949. doi:10.1016/J.GCA.2008.09.018
- [83] T. Preoanin, N. Kallay, Application of 'mass titration' to determination of surface charge of metal oxides. *Croat. Chem. Acta* 1998, 71, 1117.
- [84] S. Brunauer, P. H. Emmett, E. Teller, Adsorption of gases in multimolecular layers. J. Am. Chem. Soc. 1938, 60, 309. doi:10.1021/JA01269A023
- [85] F. Li, Y. Wang, D. Wang, F. Wei, Characterization of single-wall carbon nanotubes by N₂ adsorption. *Carbon* 2004, 42, 2375. doi:10.1016/J.CARBON.2004.02.025
- [86] B. Smith, K. Wepasnick, K. E. Schrote, H. H. Cho, W. P. Ball, D. H. Fairbrother, Influence of surface oxides on the colloidal stability of multiwalled carbon nanotubes: a structure–property relationship. *Langmuir* 2009, 25, 9767. doi:10.1021/LA901128K
- [87] R. H. Ottewill, J. N. Shaw, Electrophoretic studies on polystyrene latices. J. Electroanal. Chem. 1972, 37, 133. doi:10.1016/S0022-0728(72)80221-3
- [88] Y. P. Sun, X. Q. Li, W. X. Zhang, H. P. Wang, A method for the preparation of stable dispersion of zero-valent iron nanoparticles. *Colloids Surf.* 2007, 308, 60. doi:10.1016/J.COLSURFA. 2007.05.029
- [89] M. L. Usrey, N. Nair, D. E. Agnew, C. F. Pina, M. S. Strano, Controlling the electrophoretic mobility of single-walled carbon nanotubes: a comparison of theory and experiment. *Langmuir* 2007, 23, 7768. doi:10.1021/LA063667T
- [90] H. J. Butt, B. Cappella, M. Kappl, Force measurements with the atomic force microscope: technique, interpretation and applications. *Surf. Sci. Rep.* 2005, 59(1–6), 1. doi:10.1016/J.SURFREP.2005. 08.003
- [91] W. A. Ducker, T. J. Senden, R. M. Pashley, Direct measurement of colloidal forces using an atomic force microscope. *Nature* 1991, 353, 239. doi:10.1038/353239A0
- [92] Q. K. Ong, I. Sokojov, Attachment of nanoparticles to the AFM tips for direct measurements of interaction between a single nanoparticle and surfaces. J. Colloid Interface Sci. 2007, 310, 385. doi:10.1016/J.JCIS.2007.02.010
- [93] S. Akita, Y. Nakayama, S. Mizooka, Y. Takano, T. Okawa, Y. Miyatake, S. Yamanaka, M. Tsuji, T. Nosaka, Nanotweezers consisting of carbon nanotubes operating in an atomic force microscope. *Appl. Phys. Lett.* 2001, 79, 1691. doi:10.1063/1.1403275
- [94] H. J. Dai, J. H. Hafner, A. G. Rinzler, D. T. Colbert, R. E. Smalley, Nanotubes as nanoprobes in scanning probe microscopy. *Nature* 1996, 384, 147. doi:10.1038/384147A0

- [95] K. L. Chen, S. E. Mylon, M. Elimelech, Enhanced aggregation of alginate-coated iron oxide (hematite) nanoparticles in the presence of calcium, strontium, and barium cations. *Langmuir* 2007, 23, 5920. doi:10.1021/LA063744K
- [96] J. Rarity, Flocculation colloids stick to fractal rules. *Nature* 1989, 339, 340. doi:10.1038/339340A0
- [97] M. Y. Lin, H. M. Lindsay, D. A. Weitz, R. C. Ball, R. Klein, P. Meakin, Universality in colloid aggregation. *Nature* 1989, 339, 360. doi:10.1038/339360A0
- [98] D. A. Weitz, J. S. Huang, M. Y. Lin, J. Sung, Limits of the fractal dimension for irreversible kinetic aggregation of gold colloids. *Phys. Rev. Lett.* 1985, 54, 1416. doi:10.1103/PHYSREVLETT. 54.1416
- [99] Z. K. Zhou, P. Q. Wu, B. J. Chu, Cationic surfactant-induced fractal silica aggregates – a light-scattering study. *J. Colloid Interface* Sci. 1991, 146, 541. doi:10.1016/0021-9797(91)90218-W
- [100] R. Amal, J. A. Raper, T. D. Waite, Fractal structure of hematite aggregates. J. Colloid Interface Sci. 1990, 140, 158. doi:10.1016/ 0021-9797(90)90331-H
- [101] J. W. Zhang, J. Buffle, Multi-method determination of the fractal dimension of hematite aggregates. *Colloid. Surface A* 1996, 107, 175. doi:10.1016/0927-7757(95)03344-0
- [102] Z. K. Zhou, B. J. Chu, Light-scattering study on the fractal aggregates of polystyrene spheres – kinetic and structural approaches. J. Colloid Interface Sci. 1991, 143, 356. doi:10.1016/ 0021-9797(91)90269-E
- [103] Q. Chen, C. Saltiel, S. Manickavasagam, L. S. Schadler, R. W. Siegel, H. C. Yang, Aggregation behavior of single-walled carbon nanotubes in dilute aqueous suspension. *J. Colloid Interface Sci.* 2004, 280, 91. doi:10.1016/J.JCIS.2004.07.028
- [104] A. Y. Kim, J. C. Berg, Fractal heteroaggregation of oppositely charged colloids. *J. Colloid Interface Sci.* 2000, 229, 607. doi:10.1006/JCIS.2000.7028
- [105] J. M. López-López, A. Schmitt, A. Moncho-Jordá, R. Hidalgo-Álvarez, Stability of binary colloids: kinetic and structural aspects of heteroaggregation processes. *Soft Matter* 2006, 2, 1025. doi:10.1039/B608349H
- [106] M. A. Chappell, A. J. George, K. M. Dontsova, B. E. Porter, C. L. Price, P. Zhou, E. Morikawa, A. J. Kennedy, J. A. Steevens, Surfactive stabilization of multiwalled carbon nanotube dispersions with dissolved humic substances. *Environ. Pollut.* 2009, 157, 1081. doi:10.1016/J.ENVPOL.2008.09.039
- [107] T. Phenrat, N. Saleh, K. Sirk, R. D. Tilton, G. V. Lowry, Aggregation and sedimentation of aqueous nanoscale zero-valent iron dispersions. *Environ. Sci. Technol.* 2007, 41, 284. doi:10.1021/ES061349A
- [108] A. S. Teot, S. L. Daniels, Flocculation of negatively charged colloids by inorganic cations and anionic polyelectrolytes. *Environ. Sci. Technol.* 1969, 3, 825. doi:10.1021/ES60032A003
- [109] A. N. Giordano, H. Chaturvedi, J. C. Poler, Critical coagulation concentrations for carbon nanotubes in non-aqueous solvent. J. Phys. Chem. C 2007, 111, 11583. doi:10.1021/ JP0729866
- [110] M. Sano, J. Okamura, S. Shinkai, Colloidal nature of single-walled carbon nanotubes in electrolyte solution: the Schulze–Hardy Rule. *Langmuir* 2001, 17, 7172. doi:10.1021/LA010698+
- [111] B. Tezak, E. Matijević, K. Schulz, Coagulation of hydrophobic sols in statu nascendi. I. Determination of coagulation values. J. Phys. Chem. 1951, 55, 1557. doi:10.1021/J150492A016
- [112] H. Holthoff, S. U. Egelhaaf, M. Borkovec, P. Schurtenberger, H. Sticher, Coagulation rate measurements of colloidal particles by simultaneous static and dynamic light scattering. *Langmuir* 1996, 12, 5541. doi:10.1021/LA960326E
- [113] K. L. Chen, M. Elimelech, Aggregation and deposition kinetics of fullerene (C₆₀) nanoparticles. *Langmuir* 2006, 22, 10994. doi:10.1021/LA062072V
- [114] Y. T. He, J. Wan, T. Tokunaga, Kinetic stability of hematite nanoparticles: the effect of particle sizes. J. Nanopart. Res. 2008, 10, 321. doi:10.1007/S11051-007-9255-1

- [115] G. Sauerbrey, Verwendung von Schwingquarzen zur Wagung Dunner Schichten und zur Mikrowagung. Z. Phys. 1959, 155, 206. doi:10.1007/BF01337937
- [116] S. A. Bradford, S. R. Yates, M. Bettahar, J. Simunek, Physical factors affecting the transport and fate of colloids in saturated porous media. *Water Resour. Res.* 2002, 38, 1327. doi:10.1029/ 2002WR001340
- [117] S. A. Bradford, J. Simunek, M. Bettahar, M. T. Van Genuchten, S. R. Yates, Modeling colloid attachment, straining, and exclusion in saturated porous media. *Environ. Sci. Technol.* **2003**, *37*, 2242. doi:10.1021/ES025899U
- [118] K. L. Chen, M. Elimelech, Interaction of fullerene (C₆₀) nanoparticles with humic acid and alginate-coated silica surfaces: measurements, mechanisms, and environmental implications. *Environ. Sci. Technol.* 2008, 42, 7607. doi:10.1021/ES8012062
- [119] J. Fatisson, R. F. Domingos, K. J. Wilkinson, N. Tufenkji, Deposition of TiO₂ nanoparticles onto silica measured using a quartz crystal microbalance with dissipation monitoring. *Langmuir* 2009, 25, 6062. doi:10.1021/LA804091H
- [120] I. R. Quevedo, N. Tufenkji, Influence of solution chemistry on the deposition and detachment kinetics of a CdTe quantum dot examined using a quartz crystal microbalance. *Environ. Sci. Tech*nol. 2009, 43, 3176. doi:10.1021/ES803388U
- [121] B. L. Yuan, M. Pham, T. H. Nguyen, Deposition kinetics of bacteriophage MS2 on a silica surface coated with natural organic matter in a radial stagnation point flow cell. *Environ. Sci. Technol.* 2008, 42, 7628. doi:10.1021/ES801003S
- [122] K. M. Sirk, N. B. Saleh, T. Phenrat, H. J. Kim, B. Dufour, J. Ok, P. L. Golas, K. Matyjaszewski, G. V. Lowry, R. D. Tilton, Effect of adsorbed polyelectrolytes on nanoscale zero-valent iron particle attachment to soil surface models. *Environ. Sci. Technol.* 2009, 43, 3803. doi:10.1021/ES803589T
- [123] N. Saleh, K. Sirk, Y. Q. Liu, T. Phenrat, B. Dufour, K. Matyjaszewski, R. D. Tilton, G. V. Lowry, Surface modifications enhance nanoiron transport and NAPL targeting in saturated porous media. *Environ. Eng. Sci.* 2007, 24, 45. doi:10.1089/ EES.2007.24.45
- [124] H. W. Kroto, J. R. Heath, S. C. Obrien, R. F. Curl, R. E. Smalley, C₆₀: buckminsterfullerene. *Nature* **1985**, *318*, 162. doi:10.1038/ 318162A0
- [125] A. W. Jensen, S. R. Wilson, D. I. Schuster, Biological applications of fullerenes. *Bioorg. Med. Chem.* 1996, 4, 767. doi:10.1016/0968-0896(96)00081-8
- [126] Q. C. Ying, J. Zhang, D. H. Liang, W. Nakanishi, H. Isobe, E. Nakamura, B. Chu, Fractal behavior of functionalized fullerene aggregates. I. Aggregation of two-handed tetraaminofullerene with DNA. *Langmuir* 2005, 21, 9824. doi:10.1021/LA050557Y
- [127] R. G. Alargova, S. Deguchi, K. Tsujii, Stable colloidal dispersions of fullerenes in polar organic solvents. J. Am. Chem. Soc. 2001, 123, 10460. doi:10.1021/JA010202A
- [128] C. M. Sayes, J. D. Fortner, W. Guo, D. Lyon, A. M. Boyd, K. D. Ausman, Y. J. Tao, B. Sitharaman, et al., The differential cytotoxicity of water-soluble fullerenes. *Nano Lett.* 2004, 4, 1881. doi:10.1021/NI.0489586
- [129] J. D. Fortner, D. Y. Lyon, C. M. Sayes, A. M. Boyd, J. C. Falkner, E. M. Hotze, L. B. Alemany, Y. J. Tao, et al., C₆₀ in water: nanocrystal formation and microbial response. *Environ. Sci. Technol.* 2005, 39, 4307. doi:10.1021/ES048099N
- [130] S. J. Klaine, P. J. J. Alvarez, G. E. Batley, T. F. Fernandes, R. D. Handy, D. Y. Lyon, S. Mahendra, M. J. McLaughlin, J. R. Lead, Nanomaterials in the environment: behavior, fate, bioavailability, and effects. *Environ. Toxicol. Chem.* 2008, 27, 1825. doi:10.1897/08-090.1
- [131] R. D. Handy, F. von der Kammer, J. R. Lead, M. Hassellov, R. Owen, M. Crane, The ecotoxicology and chemistry of manufactured nanoparticles. *Ecotoxicology* 2008, 17, 287. doi:10.1007/S10646-008-0199-8
- [132] W. A. Scrivens, J. M. Tour, K. E. Creek, L. Pirisi, Synthesis of $^{14}\text{C-labeled}$ C_{60} , its suspension in water, and its uptake by human

- keratinocytes. *J. Am. Chem. Soc.* **1994**, *116*, 4517. doi:10.1021/ IA00089A067
- [133] A. Dhawan, J. S. Taurozzi, A. K. Pandey, W. Q. Shan, S. M. Miller, S. A. Hashsham, V. V. Tarabara, Stable colloidal dispersions of C₆₀ fullerenes in water: evidence for genotoxicity. *Environ. Sci. Technol.* 2006, 40, 7394. doi:10.1021/ES0609708
- [134] X. K. Cheng, A. T. Kan, M. B. Tomson, Naphthalene adsorption and desorption from aqueous C₆₀ fullerene. *J. Chem. Eng. Data* 2004, 49, 675. doi:10.1021/JE030247M
- [135] D. Bouchard, X. Ma, C. Isaacson, Colloidal properties of aqueous fullerenes: isoelectric points and aggregation kinetics of C₆₀ and C₆₀ derivatives. *Environ. Sci. Technol.* **2009**, *43*, 6597. doi:10.1021/ ES901354R
- [136] K. L. Chen, M. Elimelech, Relating colloidal stability of fullerene (C₆₀) nanoparticles to nanoparticle charge and electrokinetic properties. *Environ. Sci. Technol.* 2009, 43, 7270. doi:10.1021/ ES900185P
- [137] L. K. Duncan, J. R. Jinschek, P. J. Vikesland, C₆₀ colloid formation in aqueous systems: effects of preparation method on size, structure, and surface charge. *Environ. Sci. Technol.* 2008, 42, 173. doi:10.1021/ES071248S
- [138] J. A. Brant, J. Labille, J. Y. Bottero, M. R. Wiesner, Characterizing the impact of preparation method on fullerene cluster structure and chemistry. *Langmuir* 2006, 22, 3878. doi:10.1021/LA053293O
- [139] D. Jakubczyk, G. Derkachov, W. Bazhan, E. Lusakowska, K. Kolwas, M. Kolwas, Study of microscopic properties of water fullerene suspensions by means of resonant light scattering analysis. J. Phys. D Appl. Phys. 2004, 37, 2918. doi:10.1088/ 0022-3727/37/20/021
- [140] J. Labille, A. Masion, F. Ziarelli, J. Rose, J. Brant, F. Villieras, M. Pelletier, D. Borschneck, M. R. Wiesner, J. Y. Bottero, Hydration and dispersion of C₆₀ in aqueous systems: the nature of water– fullerene interactions. *Langmuir* 2009, 25, 11232. doi:10.1021/ LA9022807
- [141] G. V. Andrievsky, M. V. Kosevich, O. M. Vovk, V. S. Shelkovsky, L. A. Vashchenko, On the production of an aqueous colloidal solution of fullerenes. *J. Chem. Soc. Chem. Commun.* 1995, 12, 1281. doi:10.1039/C39950001281
- [142] N. O. Mchedlov-Petrossyan, V. K. Klochkov, G. V. Andrievsky, Colloidal dispersions of fullerene C₆₀ in water: some properties and regularities of coagulation by electrolytes. *J. Chem. Soc.*, *Faraday Trans.* **1997**, *93*, 4343. doi:10.1039/A705494G
- [143] S. Deguchi, R. G. Alargova, K. Tsujii, Stable dispersions of fullerenes, C₆₀ and C₇₀, in water. Preparation and characterization. *Langmuir* 2001, 17, 6013. doi:10.1021/LA010651O
- [144] J. Brant, H. Lecoanet, M. Hotze, M. Wiesner, Comparison of electrokinetic properties of colloidal fullerenes (n-C₆₀) formed using two procedures. *Environ. Sci. Technol.* 2005, 39, 6343. doi:10.1021/ES050090D
- [145] X. Ma, D. Bouchard, Formation of aqueous suspensions of fullerenes. *Environ. Sci. Technol.* 2009, 43, 330. doi:10.1021/ ES801833P
- [146] Y. G. Wang, Y. S. Li, K. D. Pennell, Influence of electrolyte species and concentration on the aggregation and transport of fullerene nanoparticles in quartz sands. *Environ. Toxicol. Chem.* 2008, 27, 1860. doi:10.1897/08-039.1
- [147] B. Espinasse, E. M. Hotze, M. R. Wiesner, Transport and retention of colloidal aggregates of C₆₀ in porous media: effects of organic macromolecules, ionic composition, and preparation method. *Environ. Sci. Technol.* 2007, 41, 7396. doi:10.1021/ES0708767
- [148] M. Terashima, S. Nagao, Solubilization of [60] fullerene in water by aquatic humic substances. *Chem. Lett.* 2007, 36, 302. doi:10.1246/ CL.2007.302
- [149] B. Xie, Z. H. Xu, W. H. Guo, Q. L. Li, Impact of natural organic matter on the physicochemical properties of aqueous C₆₀ nanoparticles. Environ. Sci. Technol. 2008, 42, 2853. doi:10.1021/ES702231G
- [150] T. E. Chang, L. R. Jensen, A. Kisliuk, R. B. Pipes, R. Pyrz, A. P. Sokolov, Microscopic mechanism of reinforcement in single-wall

- carbon nanotube/polypropylene nanocomposite. *Polymer* **2005**, *46*, 439. doi:10.1016/J.POLYMER.2004.11.030
- [151] R. H. Baughman, A. A. Zakhidov, W. A. de Heer, Carbon nanotubes – the route toward applications. *Science* 2002, 297, 787. doi:10.1126/SCIENCE.1060928
- [152] R. P. Raffaelle, B. J. Landi, J. D. Harris, S. G. Bailey, A. F. Hepp, Carbon nanotubes for power applications. *Mater. Sci. Eng. B* 2005, 116, 233. doi:10.1016/J.MSEB.2004.09.034
- [153] S. Banerjee, S. S. Wong, Rational sidewall functionalization and purification of single-walled carbon nanotubes by solution-phase ozonolysis. J. Phys. Chem. B 2002, 106, 12144. doi:10.1021/ JP026304K
- [154] Y. Peng, H. Liu, Effects of oxidation by hydrogen peroxide on the structures of multiwalled carbon nanotubes. *Ind. Eng. Chem. Res.* 2006, 45, 6483. doi:10.1021/IE0604627
- [155] I. D. Rosca, F. Watari, M. Uo, T. Akasaka, Oxidation of multiwalled carbon nanotubes by nitric acid. *Carbon* 2005, 43, 3124. doi:10.1016/J.CARBON.2005.06.019
- [156] H. Hiura, T. W. Ebbesen, K. Tanigaki, Opening and purification of carbon nanotubes in high yields. Adv. Mater. 1995, 7, 275. doi:10.1002/ADMA.19950070304
- [157] H. Hu, A. Yu, E. Kim, B. Zhao, M. E. Itkis, E. Bekyarova, R. C. Haddon, Influence of the zeta potential on the dispersability and purification of single-walled carbon nanotubes. *J. Phys. Chem. B* 2005, 109, 11520. doi:10.1021/JP050781W
- [158] N. B. Saleh, L. D. Pfefferle, M. Elimelech, Aggregation kinetics of multiwalled carbon nanotubes in aquatic systems: measurements and environmental implications. *Environ. Sci. Technol.* 2008, 42, 7963. doi:10.1021/ES801251C
- [159] H. S. Lee, C. H. Yun, Translational and rotational diffusions of multiwalled carbon nanotubes with static bending. *J. Phys. Chem.* C 2008, 112, 10653. doi:10.1021/JP803363J
- [160] D. H. Fairbrother, B. Smith, J. Wnuk, K. Wepasnick, W. P. Ball, H. Cho, F. K. Bangash, Surface oxides on carbon nanotubes (CNTs): effects on CNT stability and sorption properties in aquatic environments, in *Nanoscience and Nanotechnology, Environmental and Health Impacts* (Ed. V. Grassian) 2008, Ch. 7, pp. 131–150 (John Wiley & Sons, Inc.: New York).
- [161] A. Schierz, H. Zänker, Aqueous suspensions of carbon nanotubes: surface oxidation, colloidal stability and uranium sorption. *Environ. Pollut.* 2009, 157, 1088. doi:10.1016/J.ENVPOL.2008. 09.045
- [162] M. Cox, A. A. Pichugin, E. I. El-Shafey, Q. Appleton, Sorption of precious metals onto chemically prepared carbon from flax shive. *Hydrometallurgy* 2005, 78, 137. doi:10.1016/J.HYDROMET. 2004.12.006
- [163] M. Li, M. Boggs, T. P. Beebe, C. P. Huang, Oxidation of single-walled carbon nanotubes in dilute aqueous solutions by ozone as affected by ultrasound. *Carbon* 2008, 46, 466. doi:10.1016/J.CARBON.2007.12.012
- [164] K. Esumi, M. Ishigami, A. Nakajima, K. Sawada, H. Honda, Chemical treatment of carbon nanotubes. *Carbon* 1996, 34, 279. doi:10.1016/0008-6223(96)83349-5

- [165] Y.-T. Shieh, G.-L. Liu, H.-H. Wu, C.-C. Lee, Effects of polarity and pH on the solubility of acid-treated carbon nanotubes in different media. *Carbon* 2007, 45, 1880. doi:10.1016/J.CARBON. 2007.04.028
- [166] E. Papirer, Adsorption on Silica Surfaces 2000, Vol. 90 (Marcel Dekker, Inc.: New York, NY).
- [167] H. Hyung, J. D. Fortner, J. B. Hughes, J.-H. Kim, Natural organic matter stabilizes carbon nanotubes in the aqueous phase. *Environ. Sci. Technol.* 2007, 41, 179. doi:10.1021/ES061817G
- [168] D. Lin, B. Xing, Tannic acid adsorption and its role for stabilizing carbon nanotube suspensions. *Environ. Sci. Technol.* 2008, 42, 5917. doi:10.1021/ES800329C
- [169] H. Hyung, J.-H. Kim, Natural organic matter (NOM) adsorption to multiwalled carbon nanotubes: effect of NOM characteristics and water quality parameters. *Environ. Sci. Technol.* 2008, 42, 4416. doi:10.1021/ES702916H
- [170] X. Wang, S. Tao, B. Xing, Sorption and competition of aromatic compounds and humic acid on multiwalled carbon nanotubes. *Environ. Sci. Technol.* 2009, 43, 6214. doi:10.1021/ES901062T
- [171] K. Yang, B. S. Xing, Adsorption of fulvic acid by carbon nanotubes from water. *Environ. Pollut.* 2009, 157, 1095. doi:10.1016/ JENVPOL.2008.11.007
- [172] M. Elimelech, C. R. O'Melia, Effect of electrolyte type on the electrophoretic mobility of polystyrene latex colloids. *Colloids Surf.* 1990, 44, 165. doi:10.1016/0166-6622(90)80194-9
- [173] W. C. Hou, C. T. Jafvert, Photochemistry of aqueous C₆₀ clusters: evidence of ¹O₂ formation and its role in mediating C₆₀ phototransformation. *Environ. Sci. Technol.* 2009, 43, 5257. doi:10.1021/ ES900624S
- [174] Q. L. Li, B. Xie, Y. S. Hwang, Y. J. Xu, Kinetics of C₆₀ fullerene dispersion in water enhanced by natural organic matter and sunlight. *Environ. Sci. Technol.* 2009, 43, 3574. doi:10.1021/ES803603X
- [175] H. Hyung, J. H. Kim, Dispersion of C₆₀ in natural water and removal by conventional drinking water treatment processes. *Water Res.* 2009, 43, 2463. doi:10.1016/J.WATRES.2009.03.011
- [176] C. Y. Wang, C. Bottcher, D. W. Bahnemann, J. K. Dohrmann, A comparative study of nanometer-sized Fe(III)-doped TiO₂ photocatalysts: synthesis, characterization and activity. *J. Mater. Chem.* 2003, 13, 2322. doi:10.1039/B303716A
- [177] J. H. Choi, F. T. Nguyen, P. W. Barone, D. A. Heller, A. E. Moll, D. Patel, S. A. Boppart, M. S. Strano, Multimodal biomedical imaging with asymmetric single-walled carbon nanotube/iron oxide nanoparticle complexes. *Nano Lett.* 2007, 7, 861. doi:10.1021/ NL062306V
- [178] L. F. Shen, A. Stachowiak, S. E. K. Fateen, P. E. Laibinis, T. A. Hatton, Structure of alkanoic acid stabilized magnetic fluids. A small-angle neutron and light scattering analysis. *Langmuir* 2001, 17, 288. doi:10.1021/LA9916732

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